

Introduction to Image Processing

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@IMCF_Vinicna



github.com/vmcf-konfmi

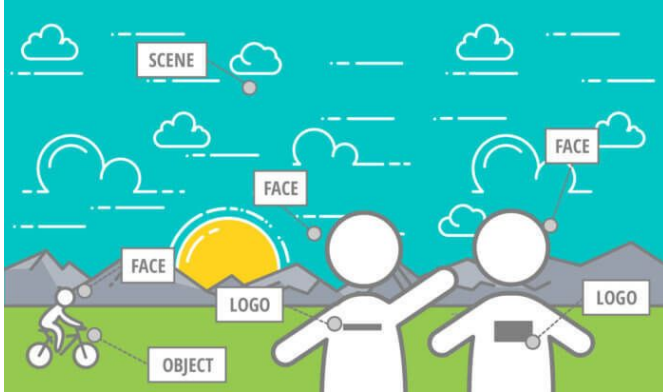


**PŘÍRODOVĚDECKÁ
FAKULTA**
Univerzita Karlova

Term Definition

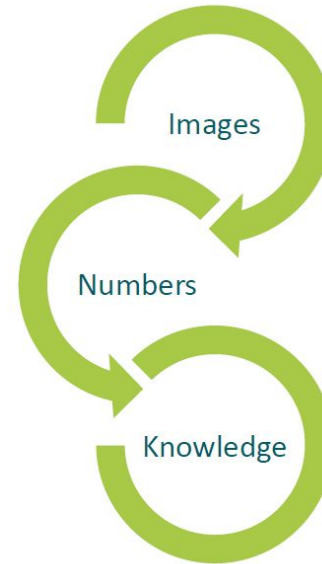
Image Analysis?

Image analysis (also known as “computer vision” or image recognition) is the ability of computers to recognize attributes within an image.



BioImage Analysis?

Understanding and quantifying microscopy & medical image data.



- Objective
- Reliable
- Reproducible
- Replicable
- Repeatable

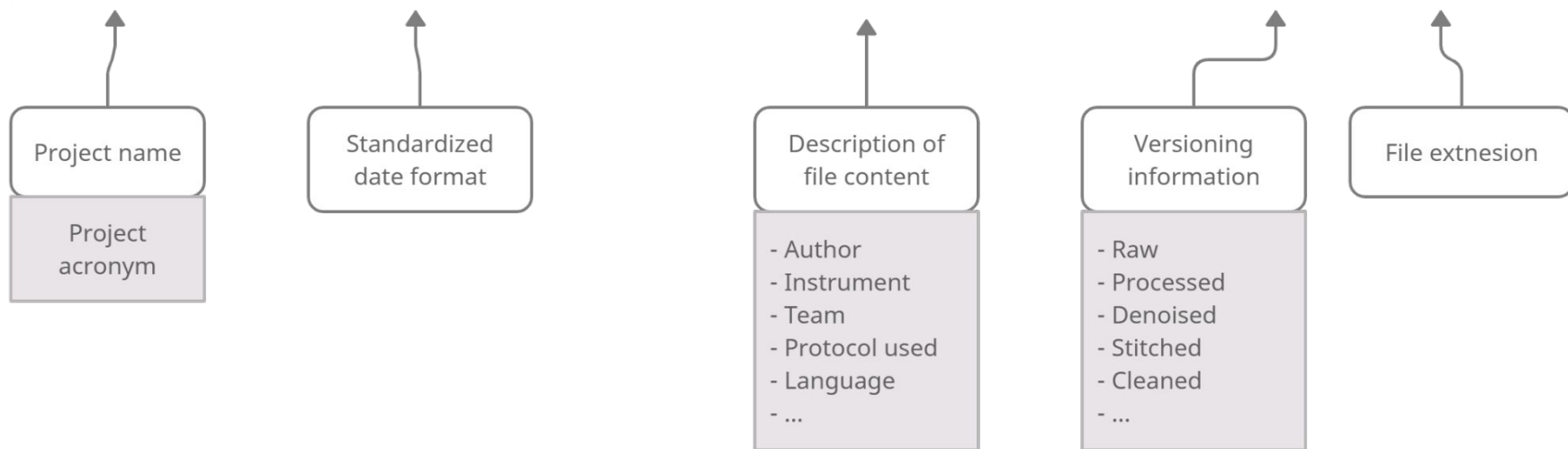
Introduction to (Bio-)Image Analysis

Bio-image analysis is supposed to be

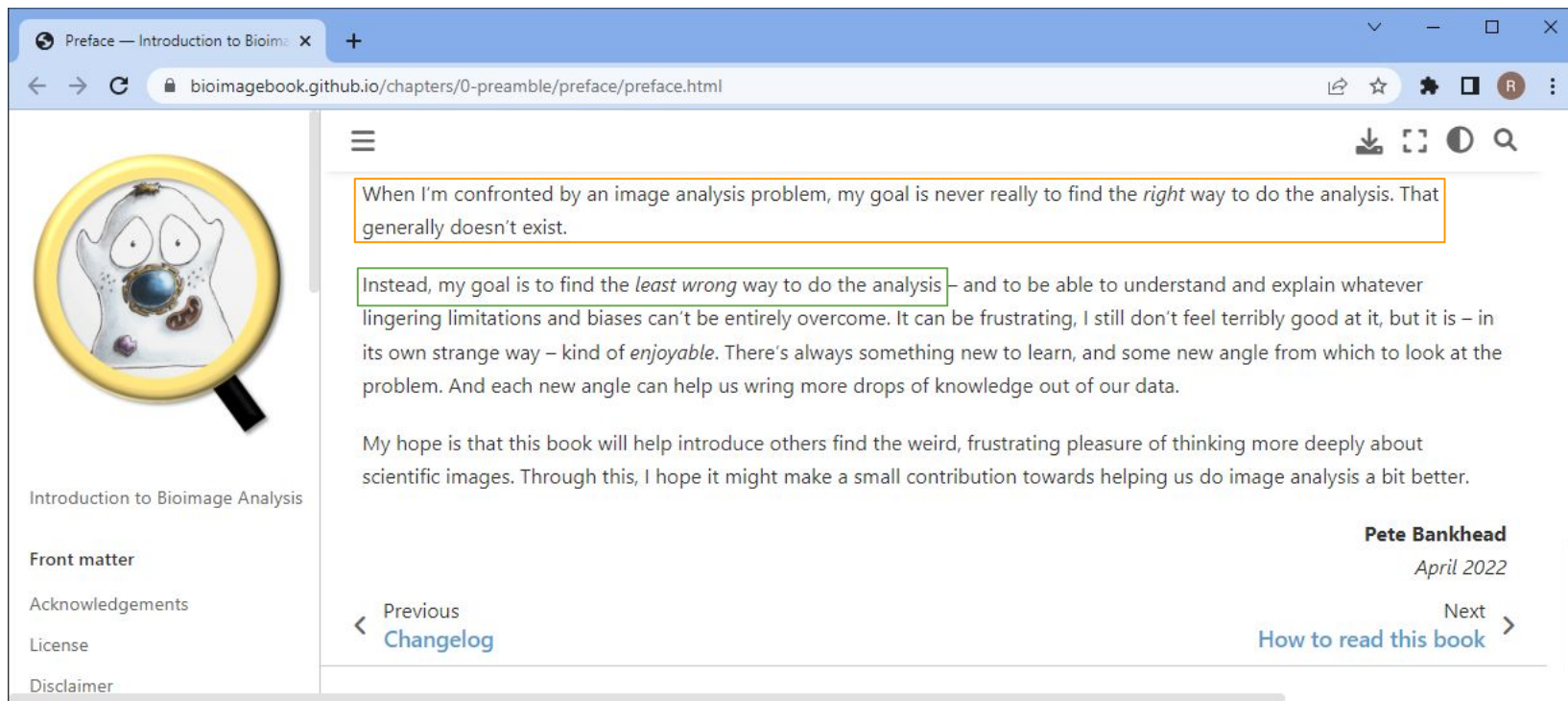
- **Quantitative**
 - We derive numbers from images which describe physical properties of the observed sample.
- **Objective**
 - The derived measurement does not depend on who did the measurement. The measurement is free of interpretation.
- **Reliable (trustworthy / validated)**
 - We are confident that the measurement is describing what it is supposed to describe.
- **Reproducible**
 - Enabling others to re-do the experiment. For this, documentation is crucial!
- **Replicability**
 - Others do execute the same analysis, potentially on other data, and see consistent results.
- **Repeatable**
 - We can do the same experiment twice under the same conditions and get the same measurements.

~~Harder~~, Better, Faster, Stronger -> Setup naming convention

Project_YYYYMMDD_ContentDescription_Version.ext




Bio-image Analysis: good scientific practice



Preface — Introduction to Bioimage Analysis

bioimagebook.github.io/chapters/0-preamble/preface/preface.html



Introduction to Bioimage Analysis

- Front matter
- Acknowledgements
- License
- Disclaimer

When I'm confronted by an image analysis problem, my goal is never really to find the *right* way to do the analysis. That generally doesn't exist.

Instead, my goal is to find the *least wrong* way to do the analysis – and to be able to understand and explain whatever lingering limitations and biases can't be entirely overcome. It can be frustrating, I still don't feel terribly good at it, but it is – in its own strange way – kind of *enjoyable*. There's always something new to learn, and some new angle from which to look at the problem. And each new angle can help us wring more drops of knowledge out of our data.

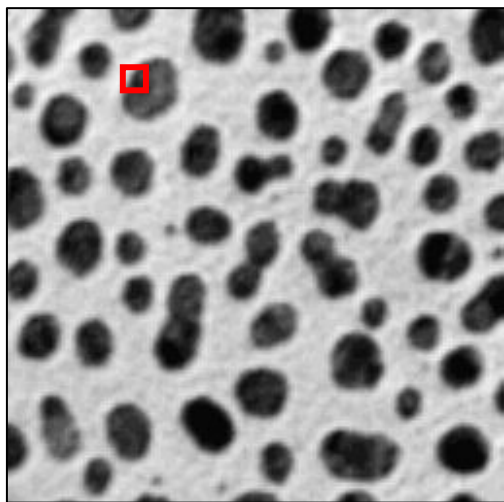
My hope is that this book will help introduce others find the weird, frustrating pleasure of thinking more deeply about scientific images. Through this, I hope it might make a small contribution towards helping us do image analysis a bit better.

Pete Bankhead
April 2022

[Previous Changelog](#) [Next How to read this book](#)

Images and pixels

- An image is just a matrix of numbers
- Pixel: “picture element”
- The edges between pixels are an artefact of the imaging / digitization. They are not real!



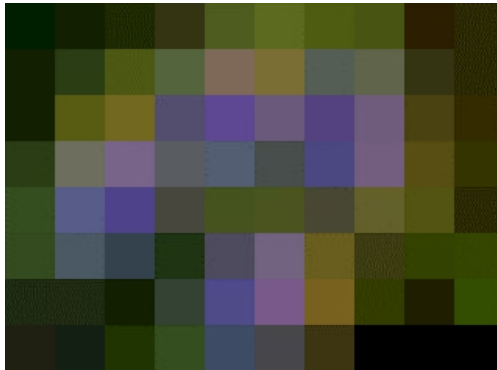
0

255

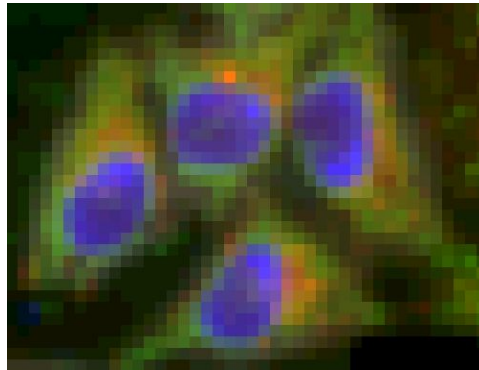
48	48	48	40	40	32	32	24	24	24	24	24	24	24
48	48	40	32	32	24	24	16	16	16	24	24	24	24
48	48	40	32	24	24	16	16	16	16	24	24	32	40
40	40	32	24	24	16	16	8	16	16	24	24	40	48
32	32	32	24	24	16	24	24	32	48	56	64	72	88
24	24	24	16	16	16	24	32	56	72	88	96	112	120
24	16	16	16	24	32	48	64	96	120	128	144	152	152
16	8	16	16	32	40	72	96	128	160	176	184	184	184
16	8	16	24	48	72	104	136	160	176	184	192	192	184
16	8	24	32	72	104	136	168	184	192	200	200	192	184
24	24	48	64	104	136	160	184	184	192	192	192	184	184
32	40	64	88	128	168	184	192	192	184	184	176	176	176
40	56	88	120	152	192	192	192	192	184	184	176	176	176
48	64	104	144	176	208	200	184	184	184	184	176	176	168

Pixel size versus resolution

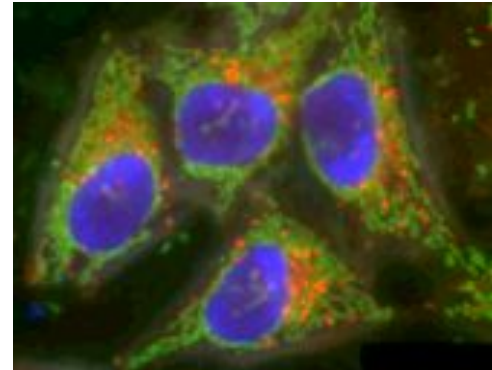
- Pixel size is a digital property of an image.
- You configure it during the imaging session at the microscope.



Pixel size: 3.3 μm



Pixel size: 0.8 μm

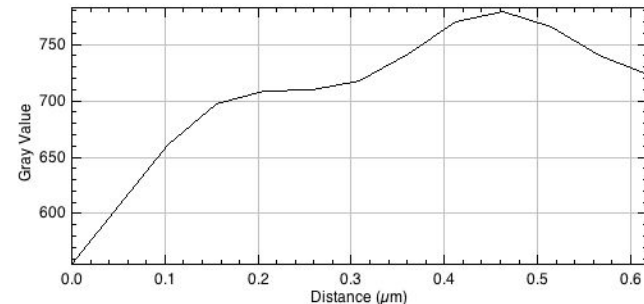
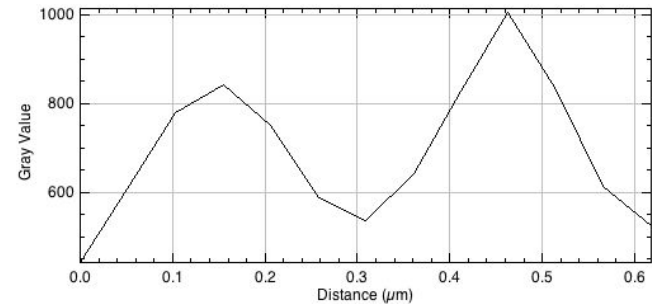
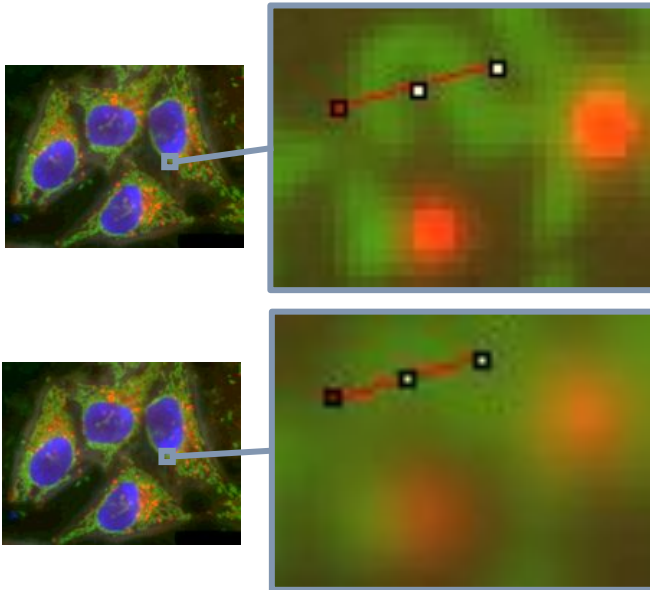


Pixel size: 0.05 μm

- We are not talking about resolution!

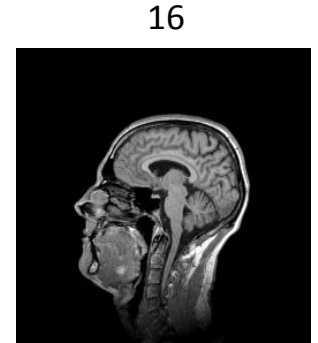
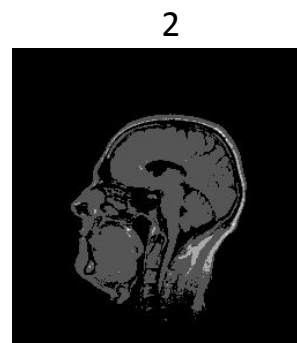
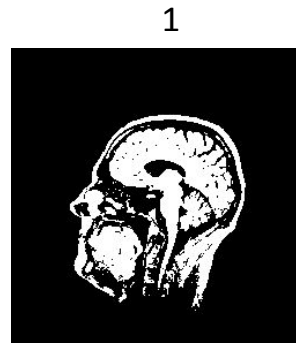
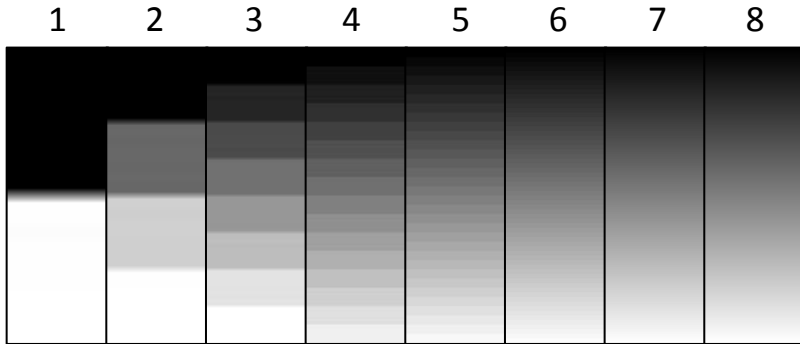
Pixel size versus resolution

- Resolution is a property of your imaging system.
- The measure of how close object can be in an image while still being differentiable, is called spatial resolution.




Bit-depth

- A bits is the smallest memory unit in computers, *atomic data*.
- The bit-depth n enumerates how many different intensity values are present in an image:
 - 2^n grey values
- In microscopy, images are usually stored as 8, 12 or 16-bit images.




Colormaps / lookup tables


- The lookup table decides how the image is displayed on screen.
- Applying a different lookup table does not change the image. All pixel values stay the same, they just appear differently

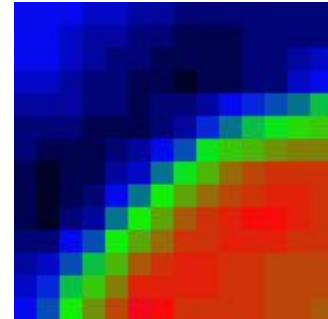
Pixel value	Display
0	
1	
2	
...	
255	



Pixel value	Display
0	
1	
2	
...	
255	



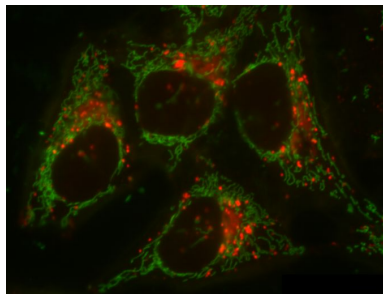
Pixel value	Display color
0	
1	
2	
...	
255	



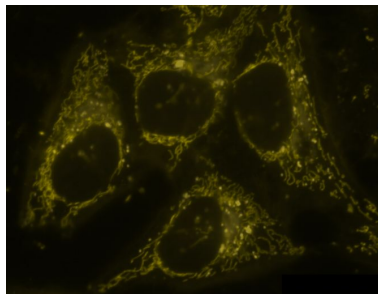
Colormaps / lookup tables

- Choose visualization of your color tables wisely!
- Think of people with red/green blindness!

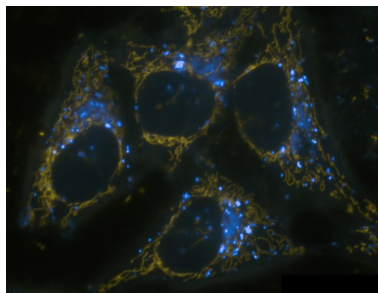
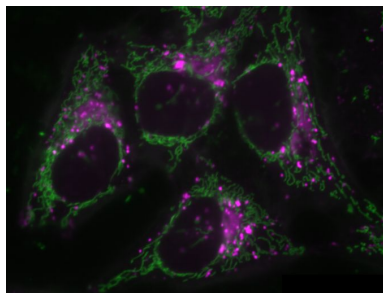
Common view



Red/green blind people may see it like this



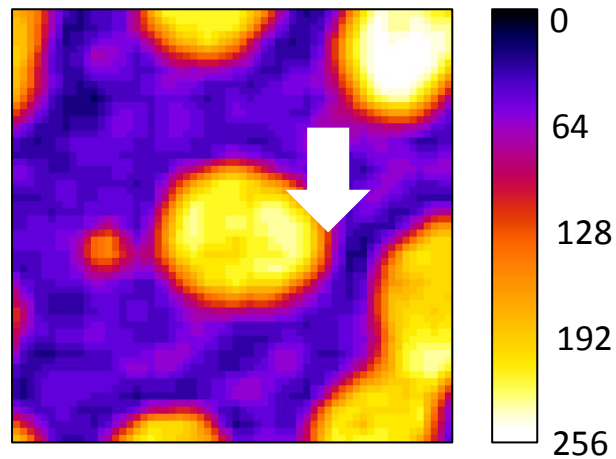
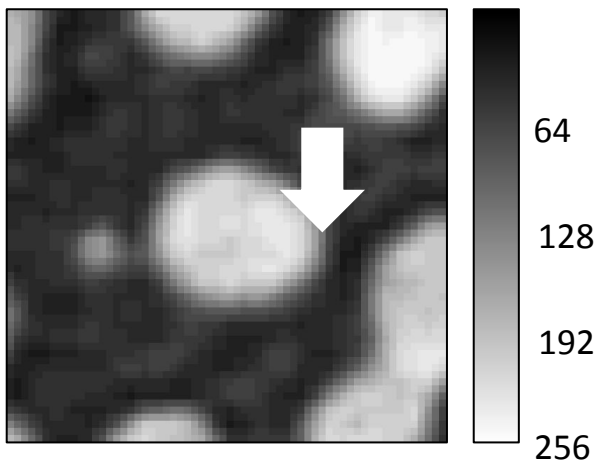
Replace red
with
magenta!



Colormaps / lookup tables

Which intensity does the marked pixel have?

0



0

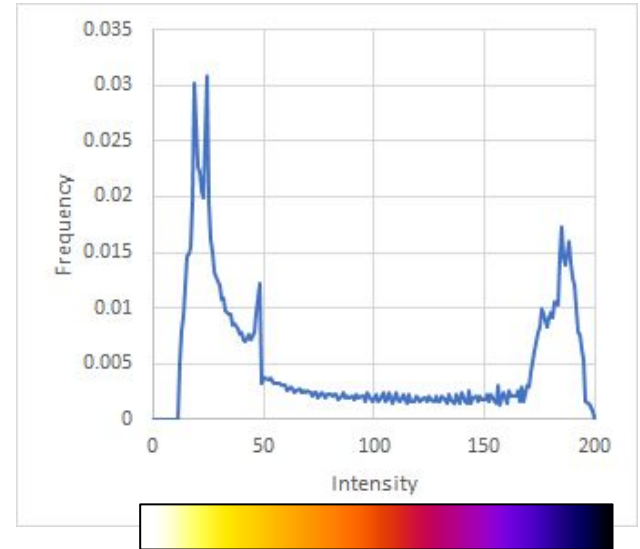
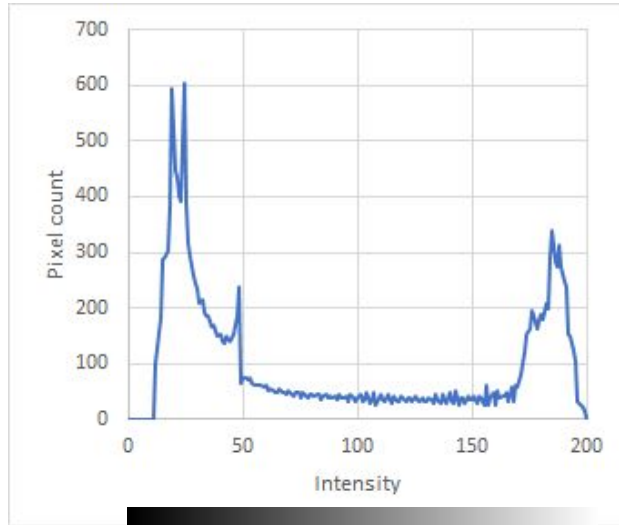
64

128

192

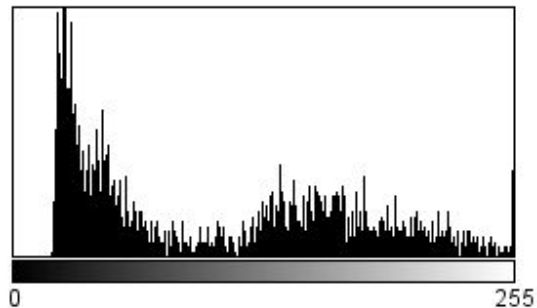
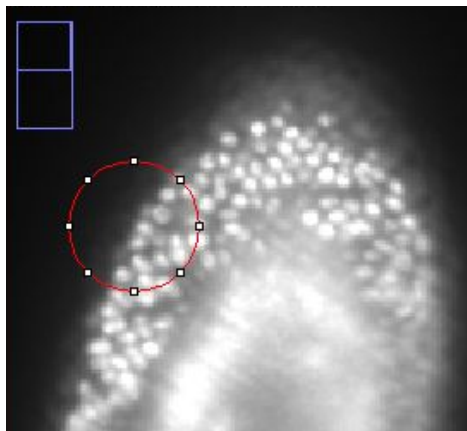
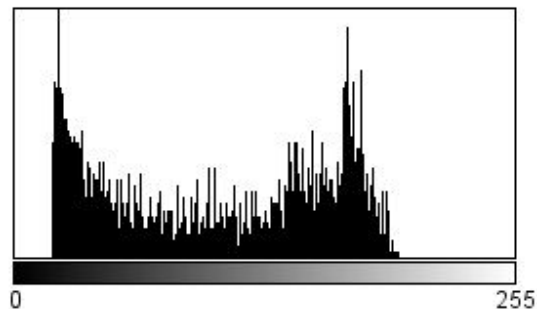
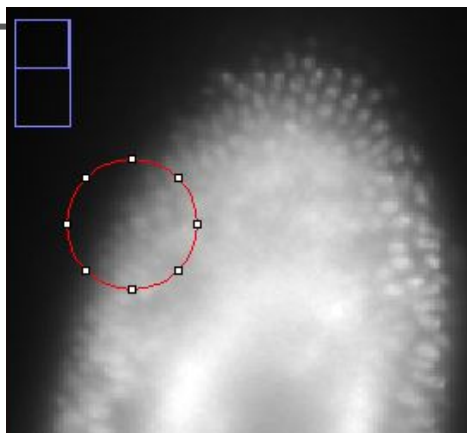
Histograms

- A histogram shows the probability distribution of pixel intensities.
- The probability of a pixel having a certain grey value can be measured by counting pixels and calculating the frequency of the given intensity.
- Whenever you see a histogram, try to imagine the lookup-table on the X-axis



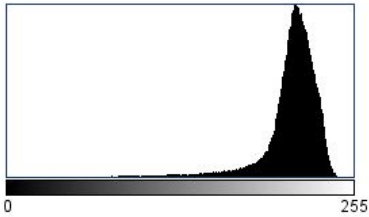
Histograms

- Histograms are summaries of images
- Tell stories, e.g. about image quality

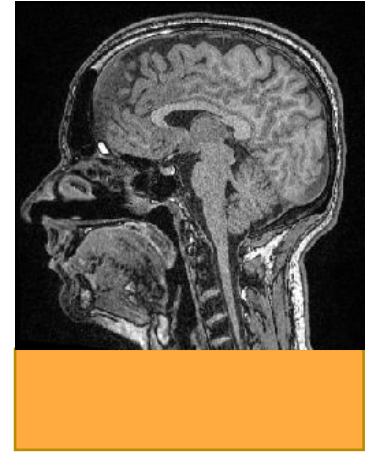
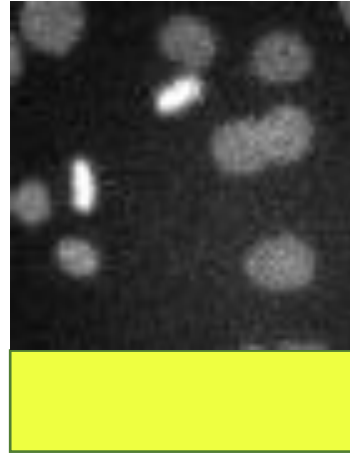


Histograms

To which of the three images does this histogram belong to?

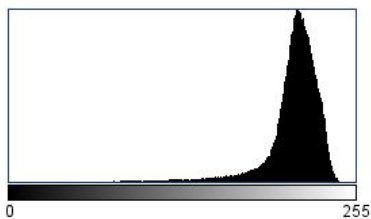


N: 165648	Min: 1
Mean: 207.819	Max: 253
StdDev: 25.834	Mode: 212 (5234)
Value: 200	Count: 2219

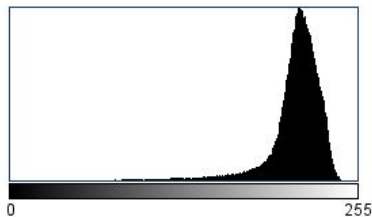


Histograms

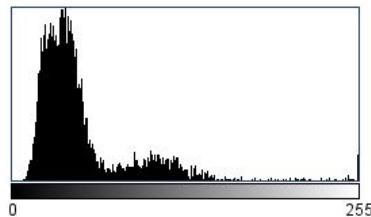
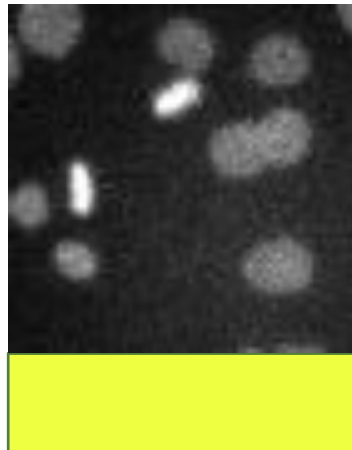
To which of the three images does this histogram belong to?



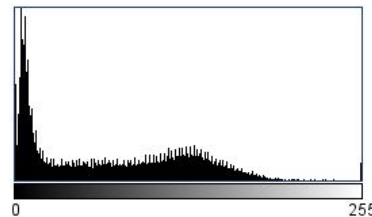
N: 165648
Mean: 207.819
StdDev: 25.834
Value: 200
Min: 1
Max: 253
Mode: 212 (5234)
Count: 2219



N: 165648
Mean: 207.819
StdDev: 25.834
Value: 200
Min: 1
Max: 253
Mode: 212 (5234)
Count: 2219



N: 4900
Mean: 51.060
StdDev: 36.426
Value: 64
Min: 8
Max: 255
Mode: 39 (125)
Count: 9



N: 65536
Mean: 70.929
StdDev: 59.567
Value: 59
Min: 0
Max: 255
Mode: 4 (2352)
Count: 239

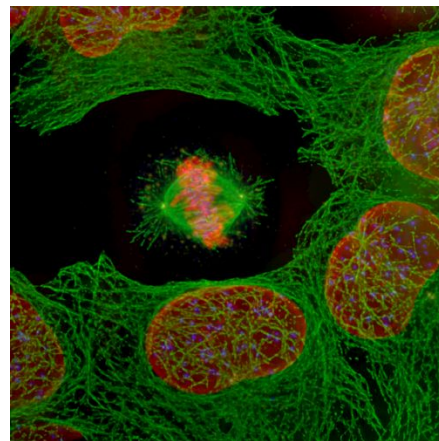
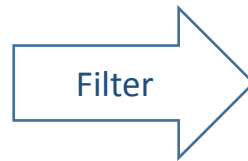
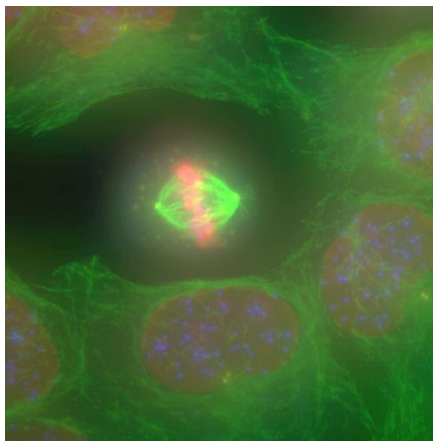
Image Processing and Filtering

Filters

- An image processing filter is an operation on an image.
- It takes an image and produces a new image out of it.
- Filters change pixel values.

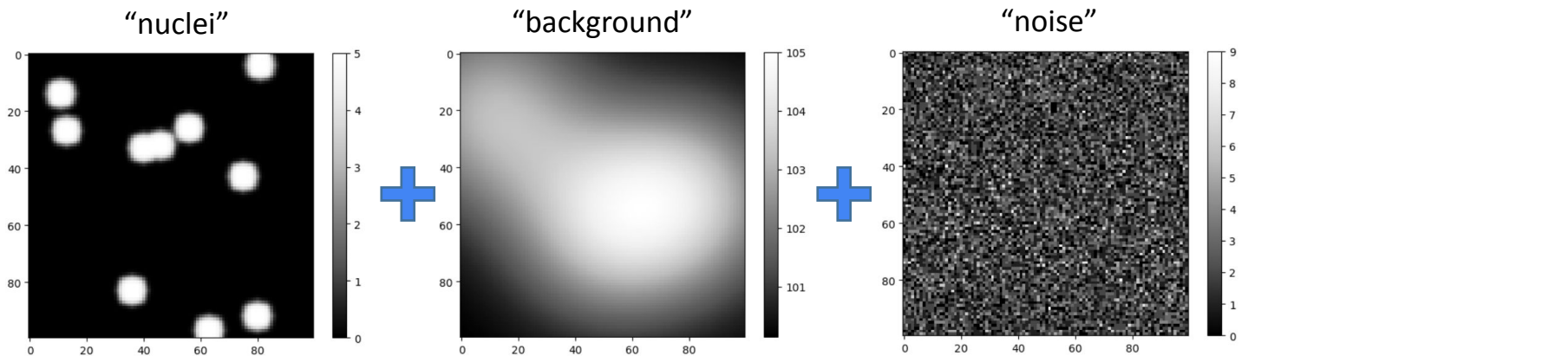
- There is no “best” filter. Which filter fits your needs, depends on the context.
- Filters do not do magic. They can not make things visible which are not in the image.

- Application examples
 - Noise-reduction
 - Artefact-removal
 - Contrast enhancement
 - Correct uneven illumination



Effects harming image quality

- Image formation (simulated)



- Aberrations, defocus
- Motion blur

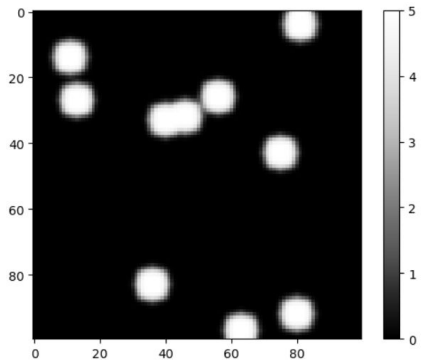
- Light from objects behind and in front of the scene (out-of-focus light)
- Dirt on the object slide
- Camera offset

- Shot noise (arriving photons)
- Dark noise (electrons made from photons)
- Read-out-noise (electronics)

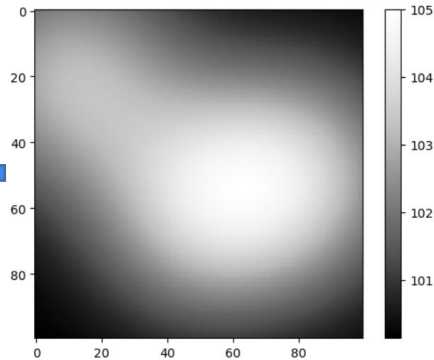
Effects harming image quality

- Image formation (simulated)

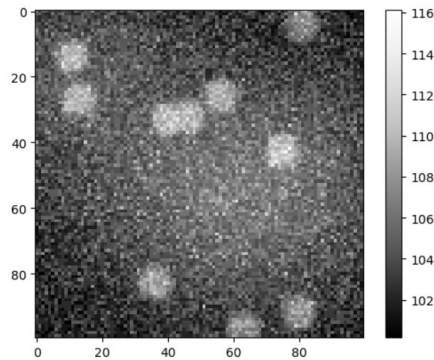
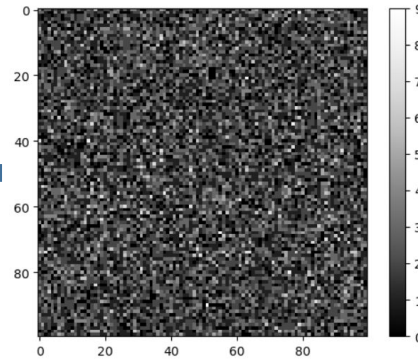
“nuclei”



“background”



“noise”



Segmentation

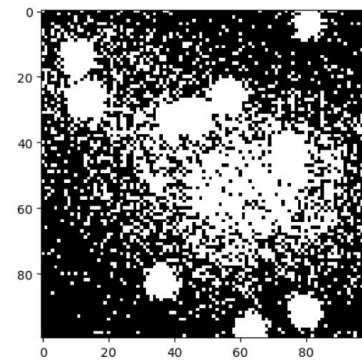


Image filtering

- We need to remove the noise to help the computer *interpreting* the image

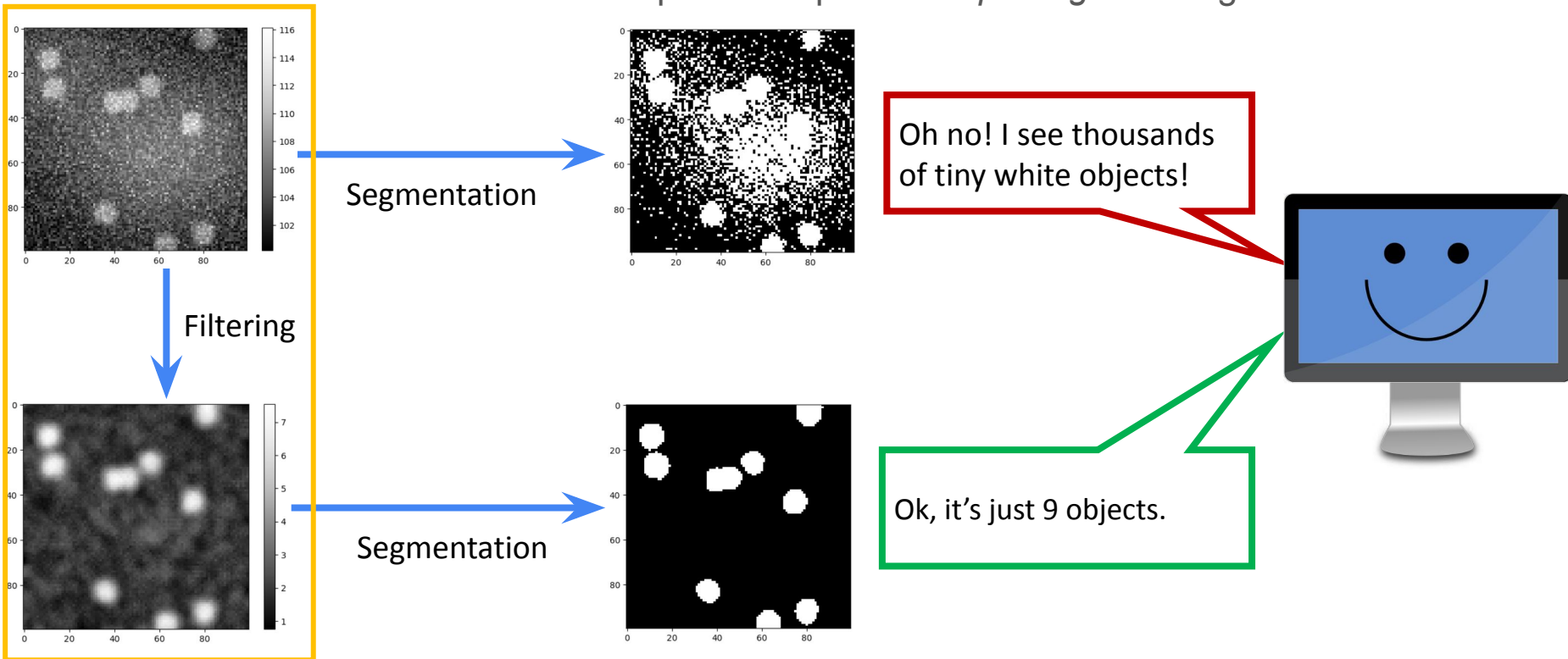


Image filtering

- Attempt to invert / “undo” processes disturbing image quality

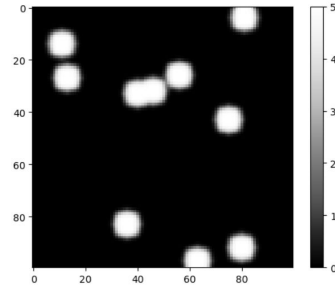
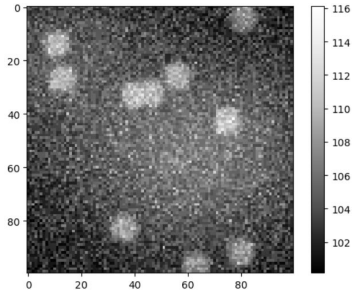
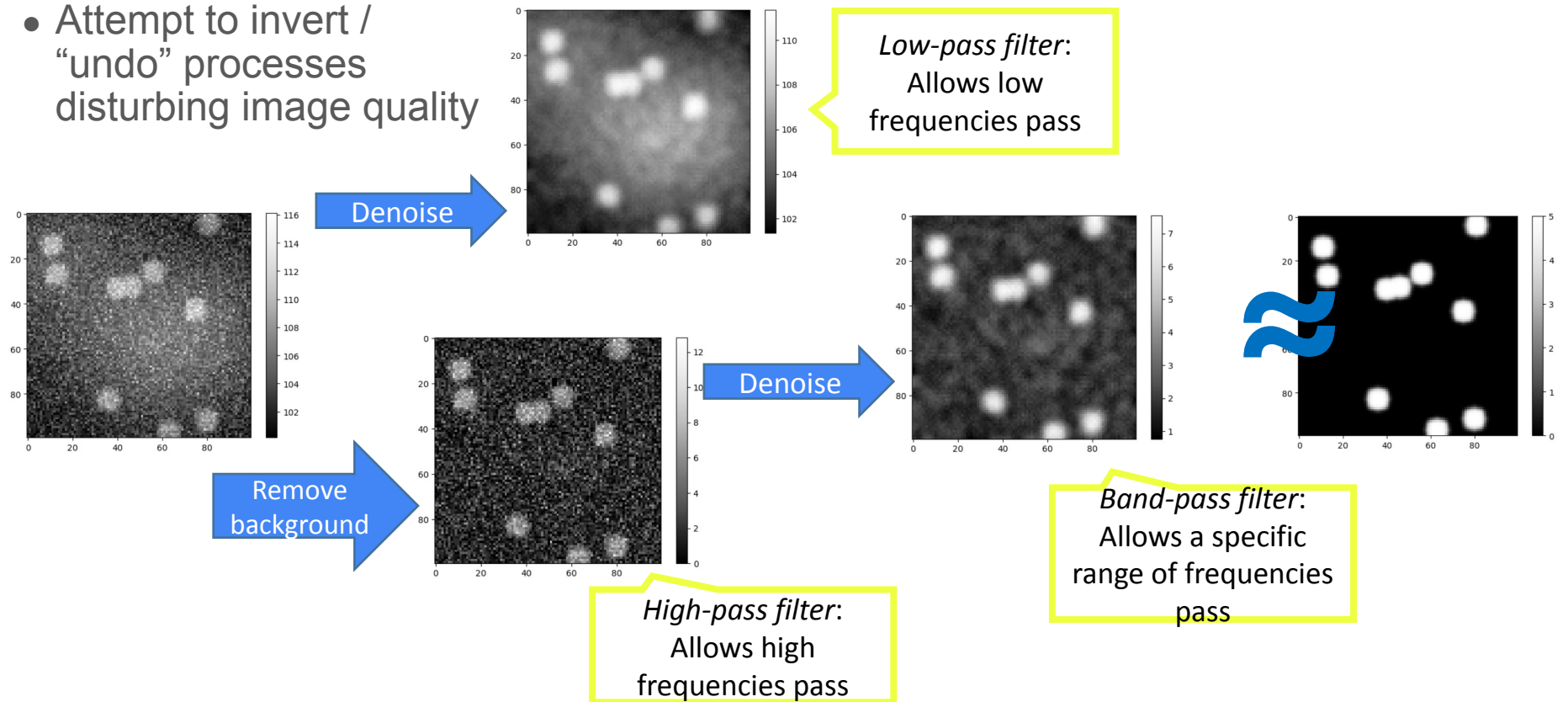


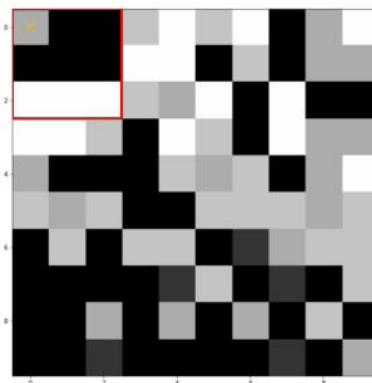
Image filtering

- Attempt to invert / “undo” processes disturbing image quality



Linear Filters

- *Linear filters* replace each pixel value with a weighted linear combination of surrounding pixels
- Filter *kernels* are matrices describing a linear filter
- This multiplication of surrounding pixels according to a matrix is called *convolution*



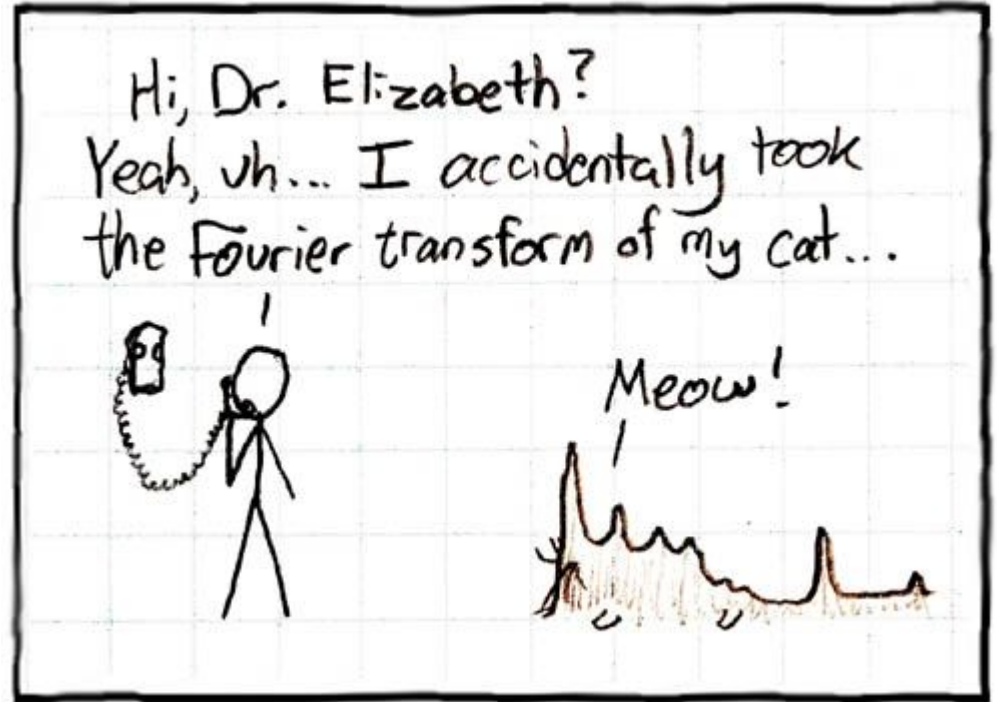
Mean filter, 3x3 kernel

$$\begin{bmatrix} 1/9 & 1/9 & 1/9 \\ 1/9 & 1/9 & 1/9 \\ 1/9 & 1/9 & 1/9 \end{bmatrix}$$

Linear Filters

Also we can apply them with
Fourier transform

But... let's skip that for "safety
reasons". And try to stick with
using convolution in spatial
domain.



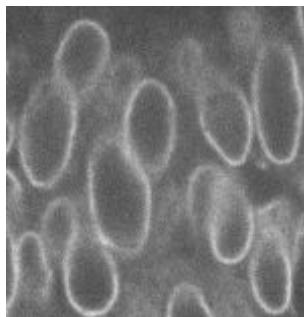
Linear filters

- Terminology:

- “We convolve an image with a kernel.”
- Convolution operator: *

- Examples

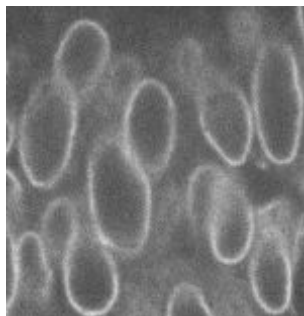
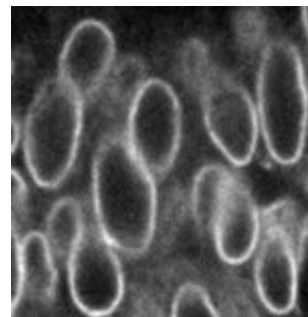
- Mean
- Gaussian blur
- Sobel-operator
- Laplace-filter



*

$$\begin{bmatrix} 1 & 1 & 1 \\ 1 & 8 & 1 \\ 1 & 1 & 1 \end{bmatrix}$$

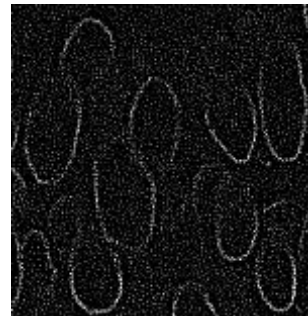
=



*

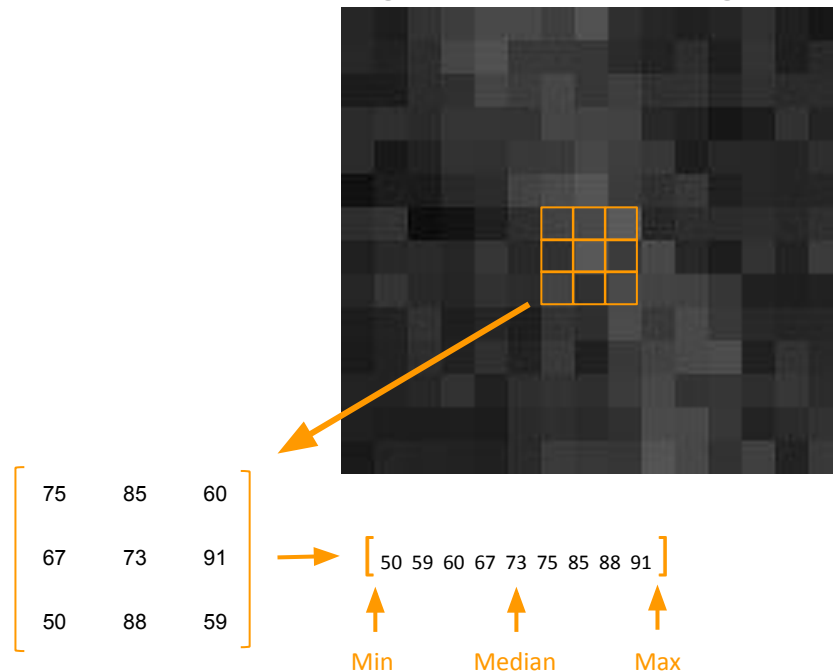
$$\begin{bmatrix} 0 & -1 & 0 \\ -1 & 4 & -1 \\ 0 & -1 & 0 \end{bmatrix}$$

=



Nonlinear Filters

- Non linear filters also replace pixel value inside as rolling window but using a non-linear function.
- Examples: order statistics filters
 - Min
 - Median
 - Max
 - Variance
 - Standard deviation



Noise removal

- Gaussian filter
- Median filter (computationally expensive)

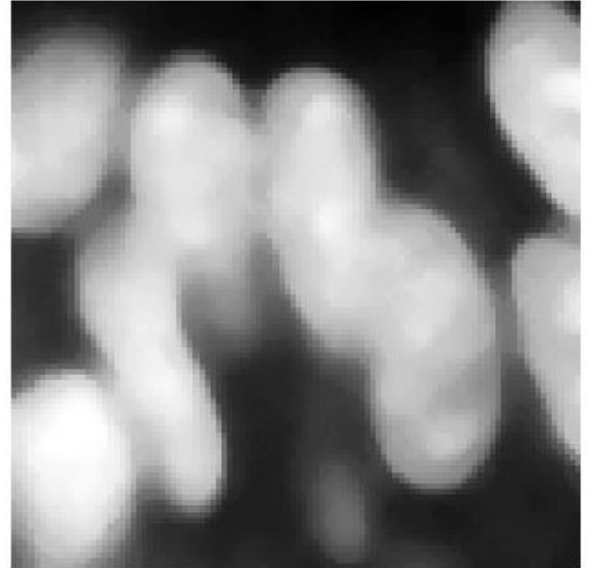
Original



Gaussian

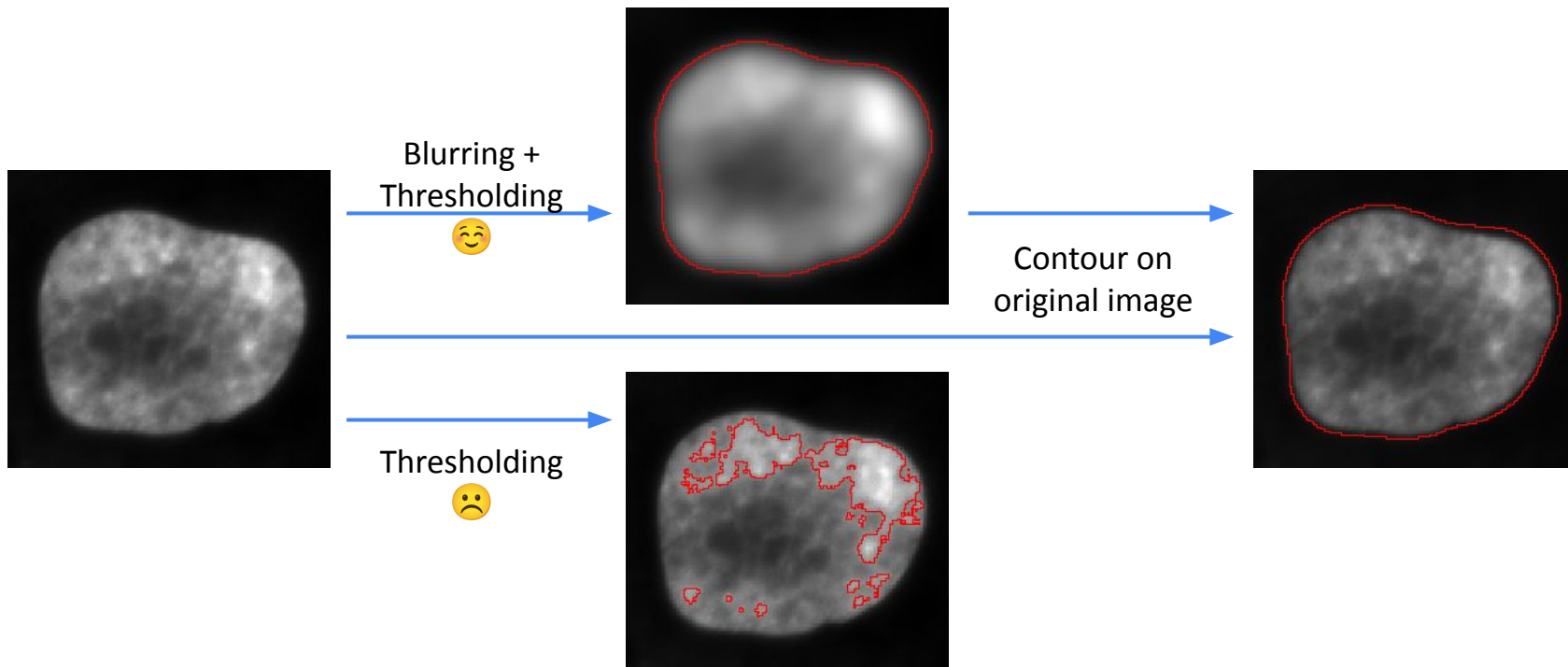


Median



Filtering for improving thresholding results

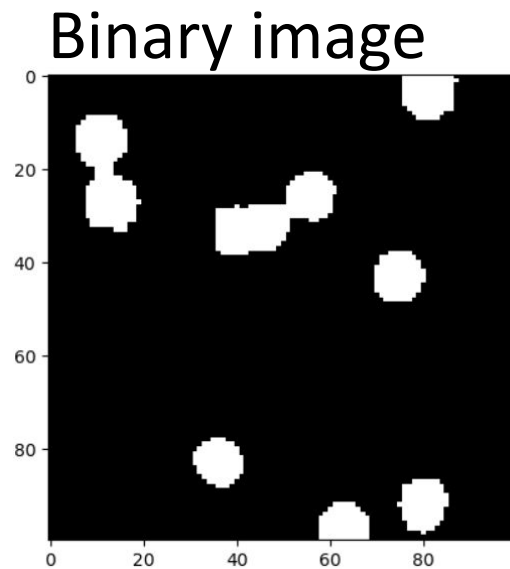
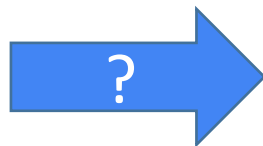
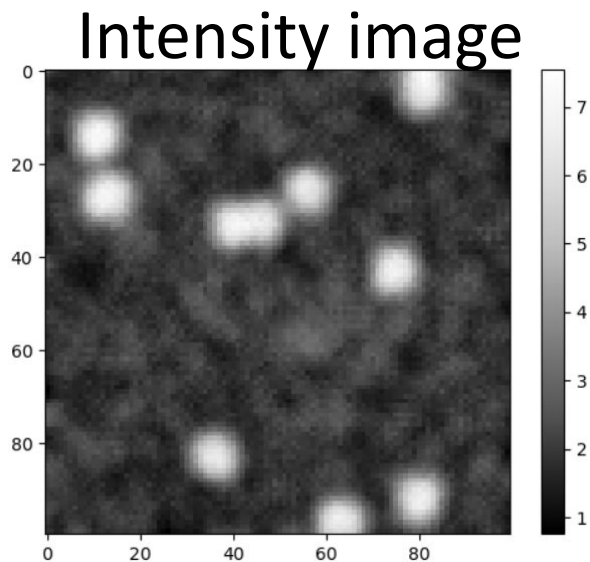
- In case thresholding algorithms outline the wrong structure, blurring in advance may help.
- However: **Do not** continue processing the blurred image, continue with the original!



Segmentation / binarization

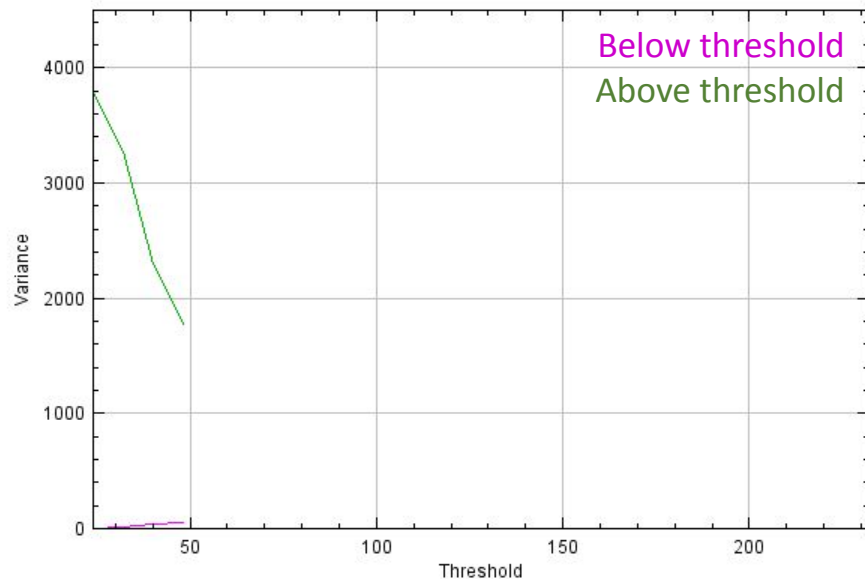
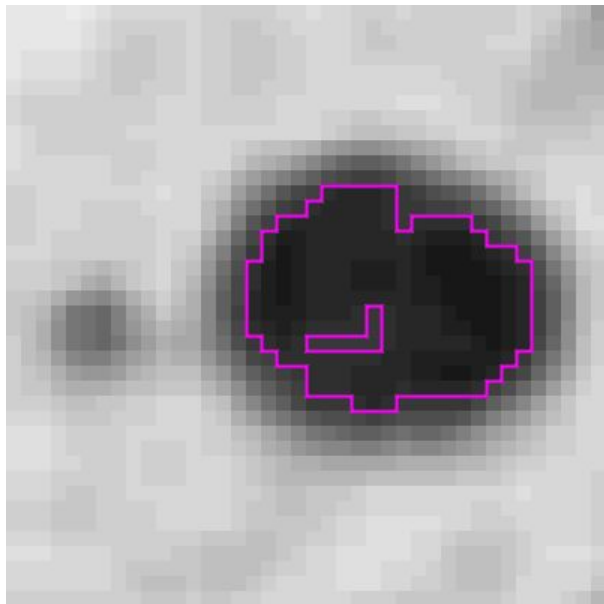
Thresholding

- Very basic and yet efficient segmentation technique
- Histogram based, to determine an intensity threshold
- Not state-of-the-art in many fields (anymore)



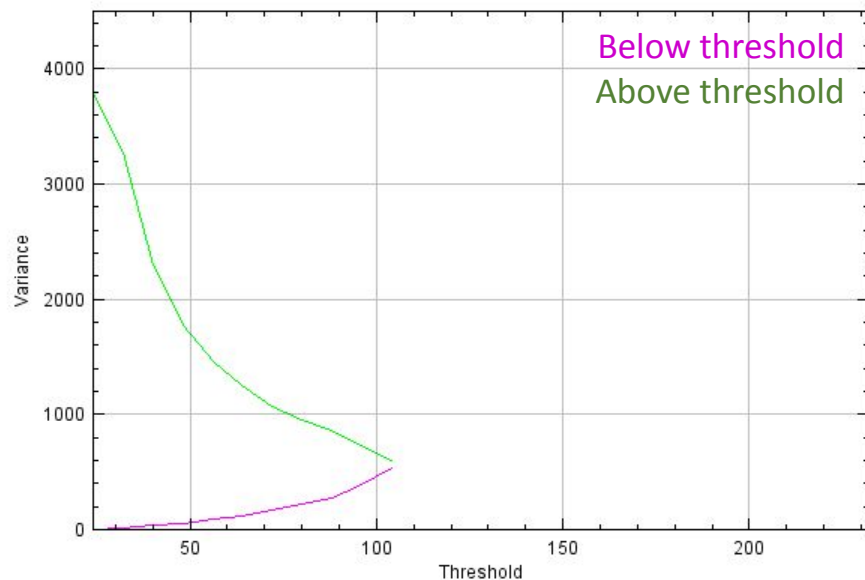
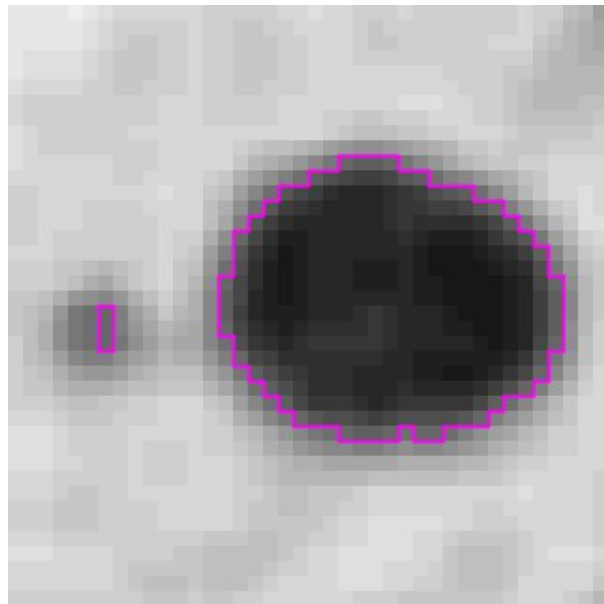
Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.



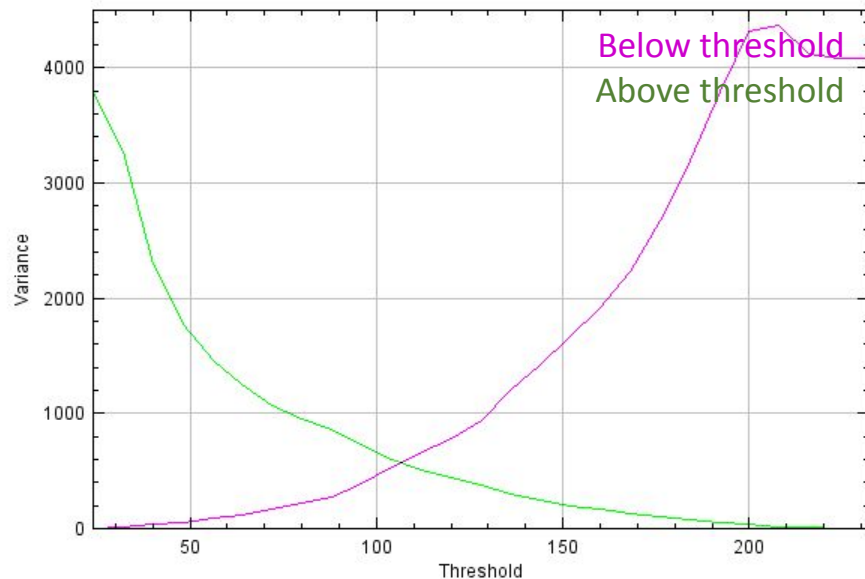
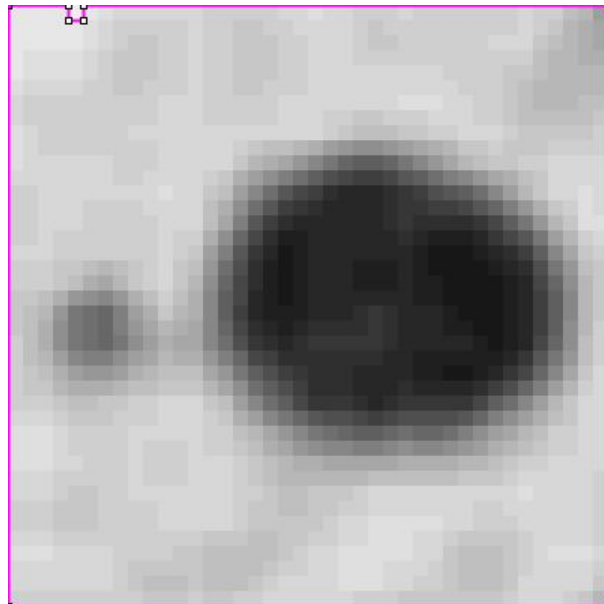
Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.



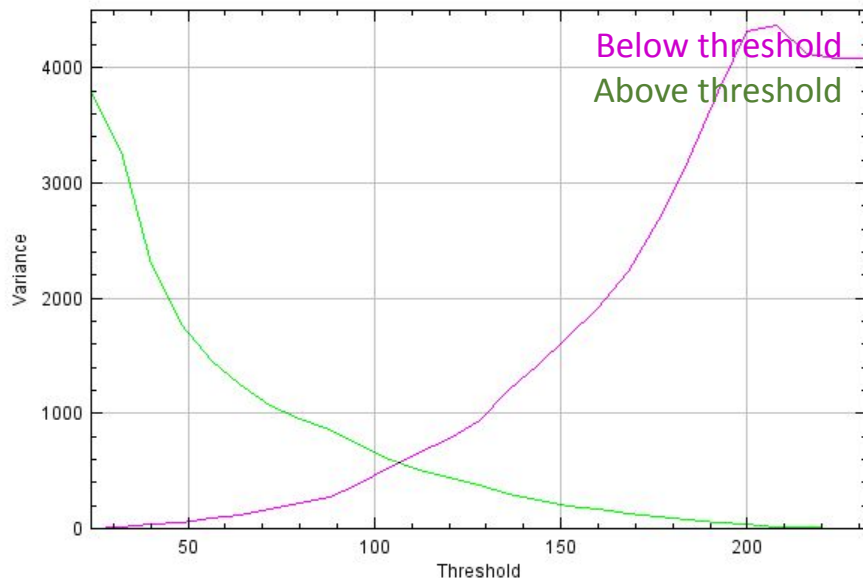
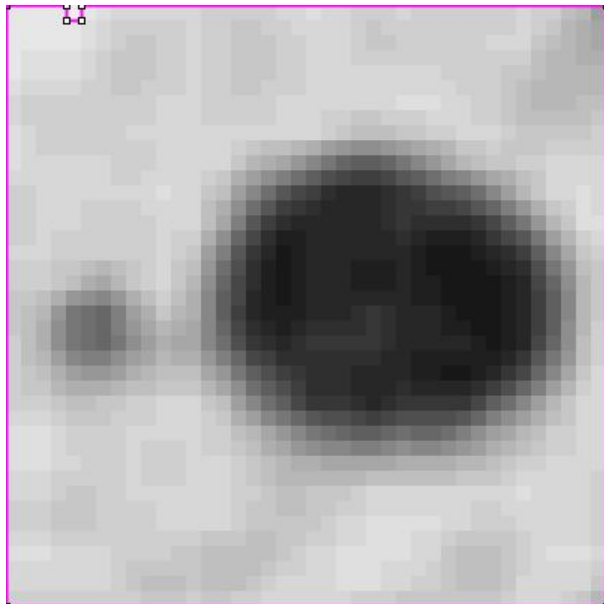
Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.



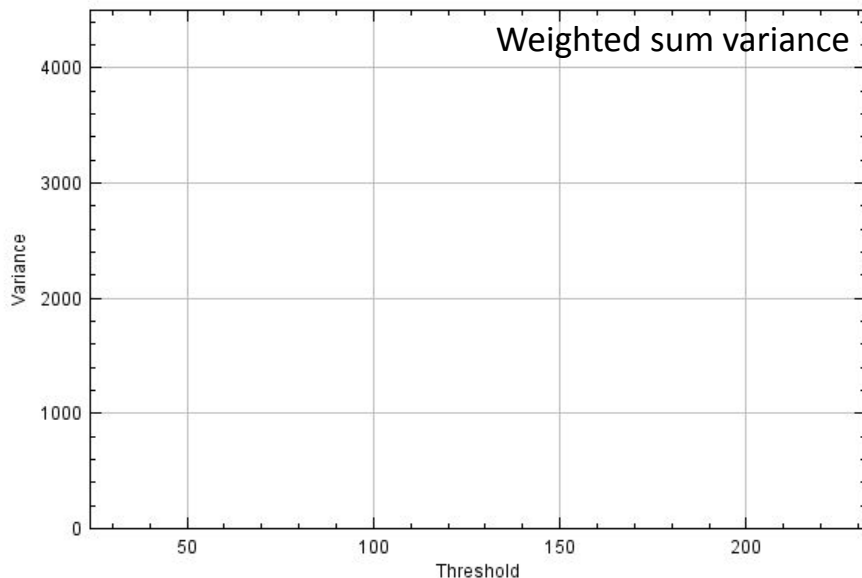
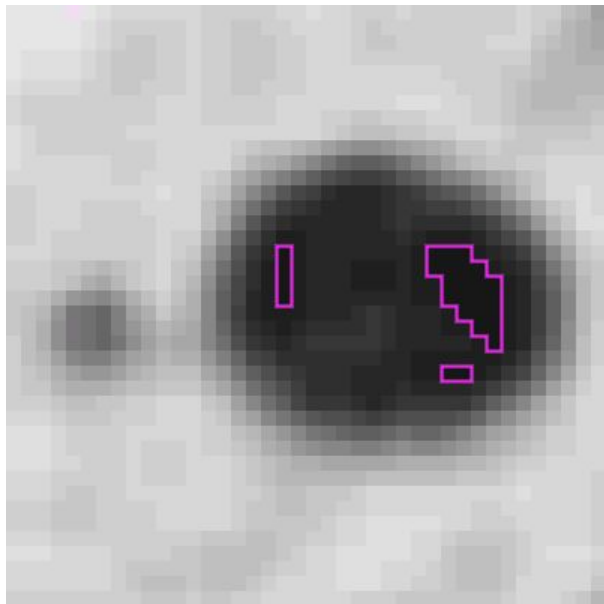
Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.
- Weighted (!) sum variance



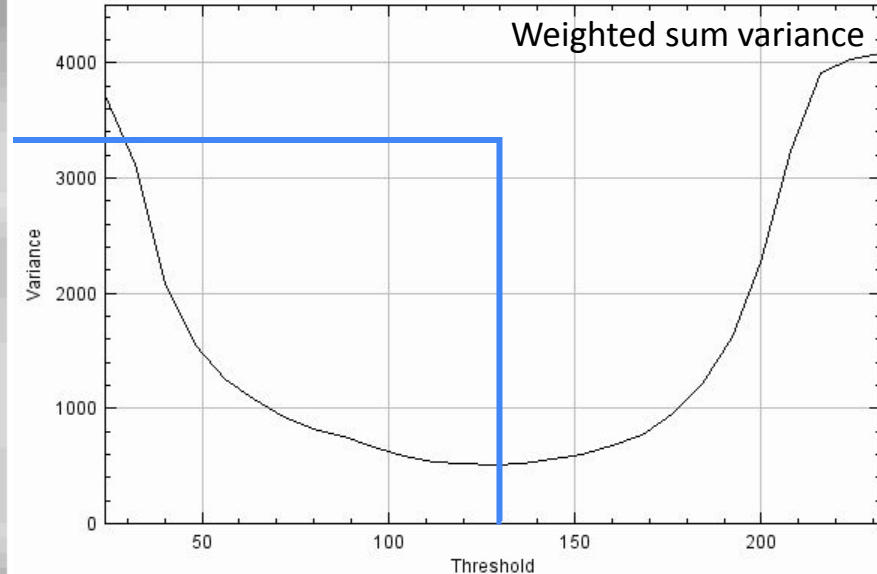
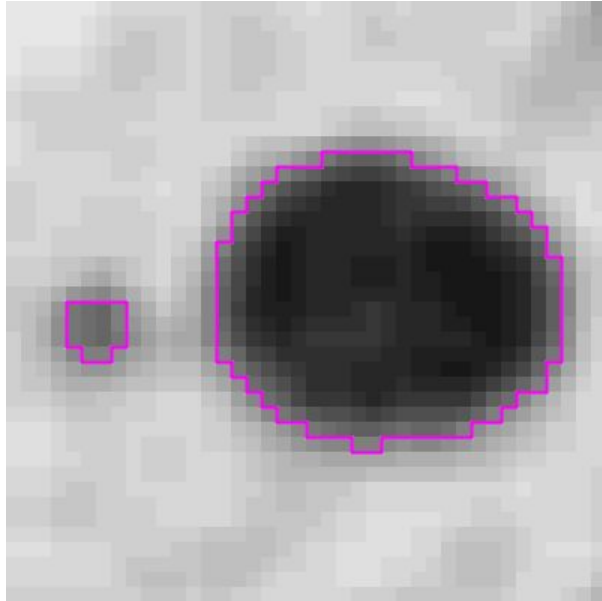
Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.
- Weighted (!) sum variance



Thresholding: Otsu's method

- Searching for a threshold where the variance in both classes (above/below threshold) becomes minimal.
- Weighted (!) sum variance

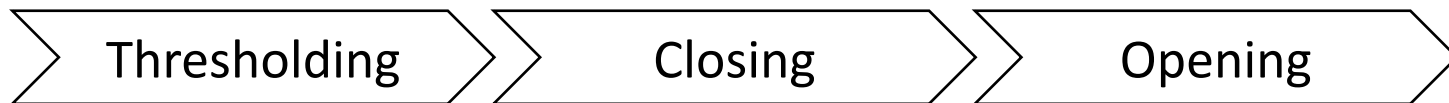


See also: <http://www.labbookpages.co.uk/software/imgProc/otsuThreshold.html>

Morphological operations

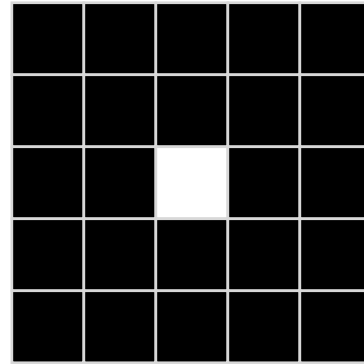
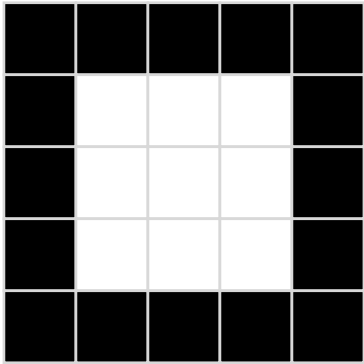
Refining masks

- Binary mask images may not be perfect immediately after thresholding.
- There are ways of refining them



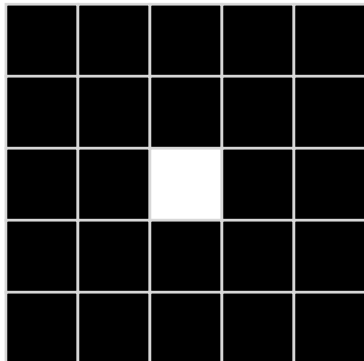
Erosion

- Erosion: Every pixel with at least one black neighbor becomes black.



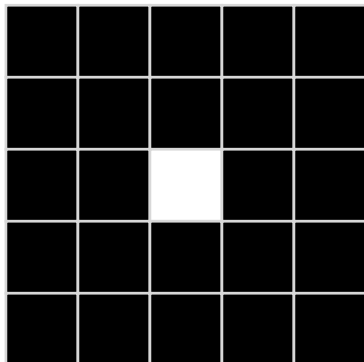
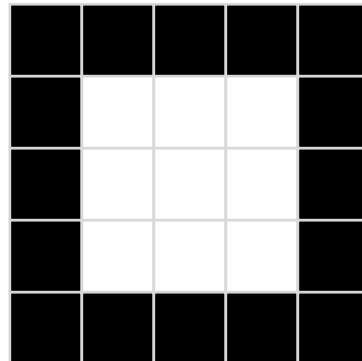
Dilation

- Dilation: Every pixel with at least one white neighbor becomes white.



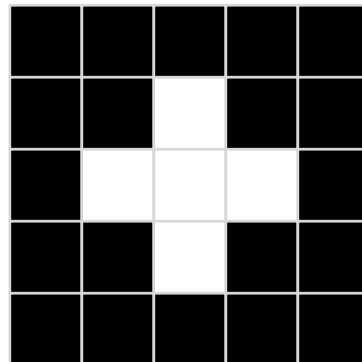
Dilation

8-connected neighborhood



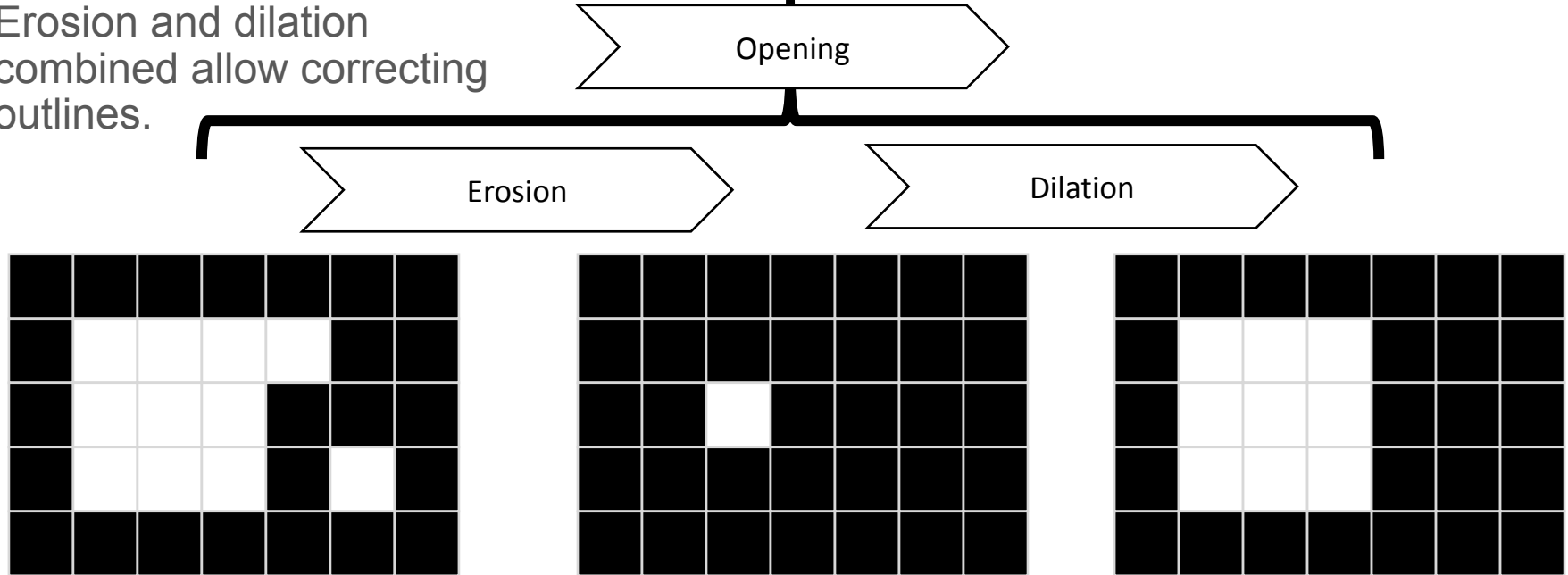
Dilation

4-connected neighborhood



Opening

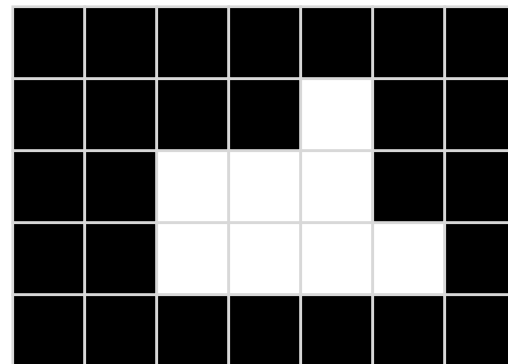
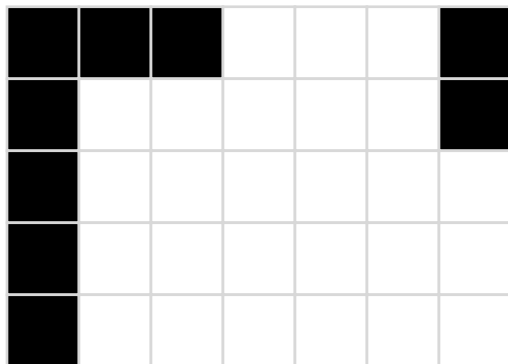
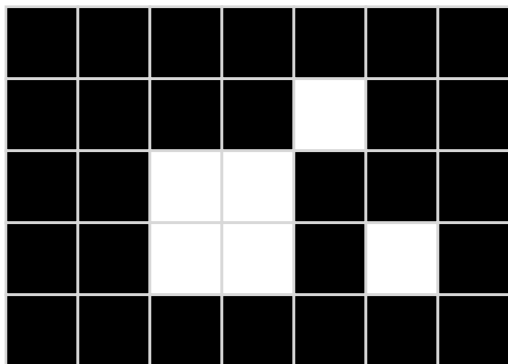
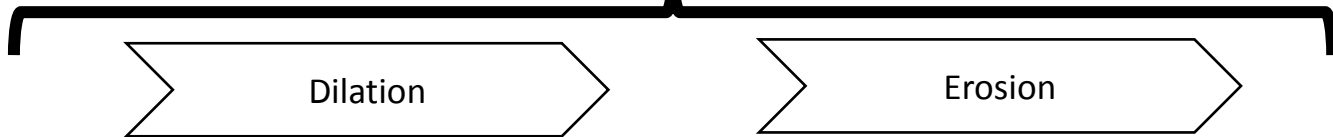
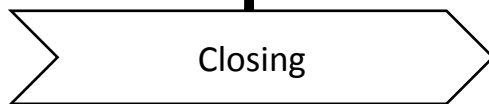
Erosion and dilation combined allow correcting outlines.



- It can separate white (high intensity) structures that are weakly connected
- It may erase small white structures

Closing

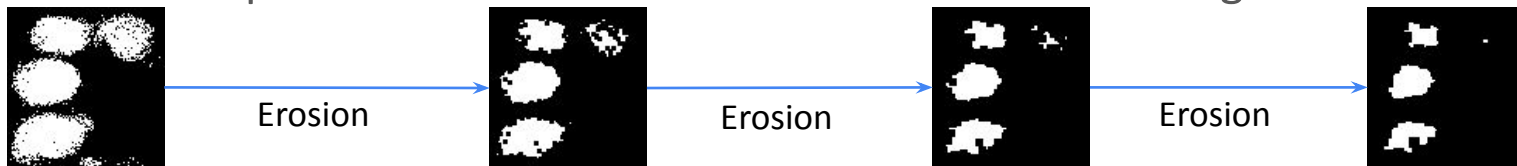
Erosion and dilation combined allow correcting outlines.



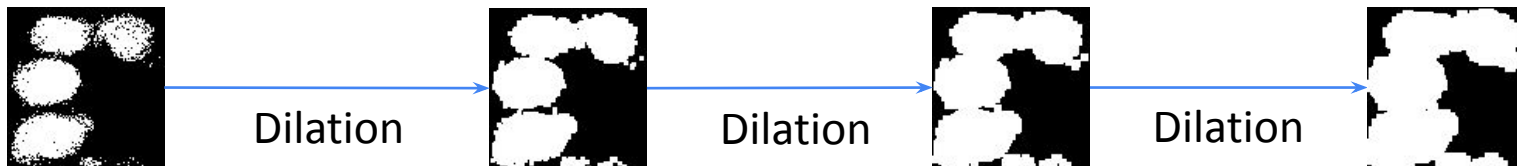
- It can connect white (high intensity) structures that are nearby
- It may close small holes inside structures

Chaining erosion and dilation

Erosion: Set all pixels to black which have at least one black neighbor.



Dilation: Set all pixels to white which have at least one white neighbor.



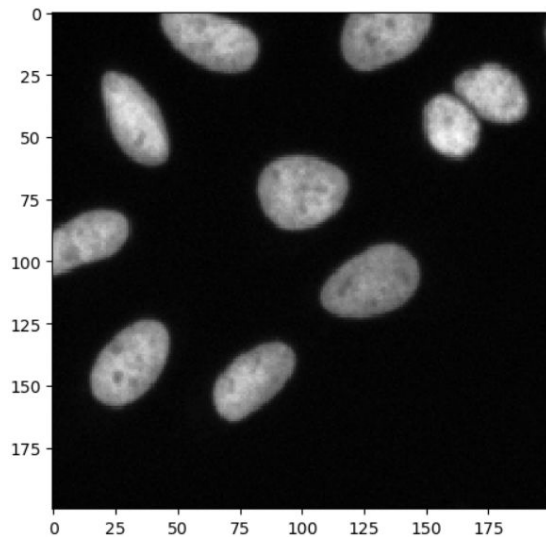
Closing: Dilation + Erosion



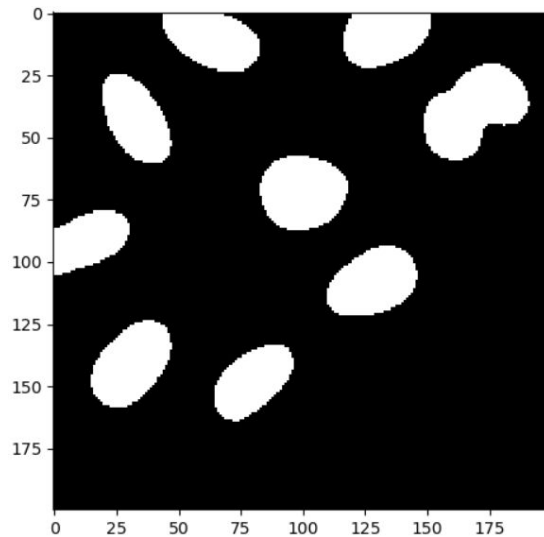
Opening: Erosion + Dilation

Terminology

Intensity image



Binary image

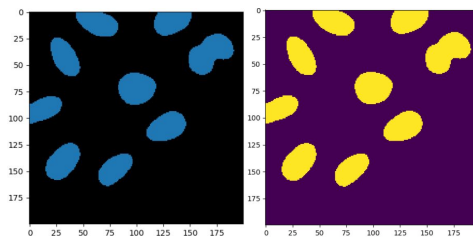


Label image



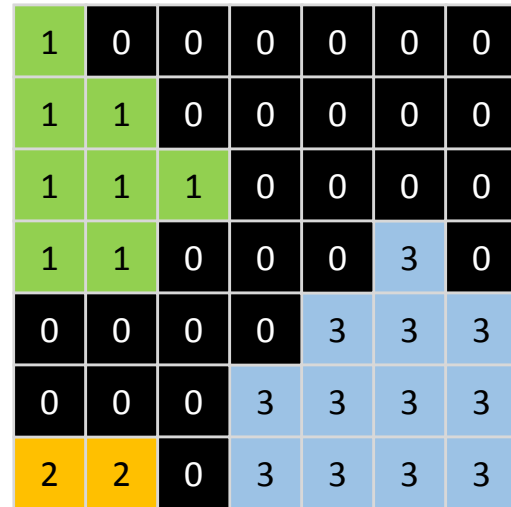
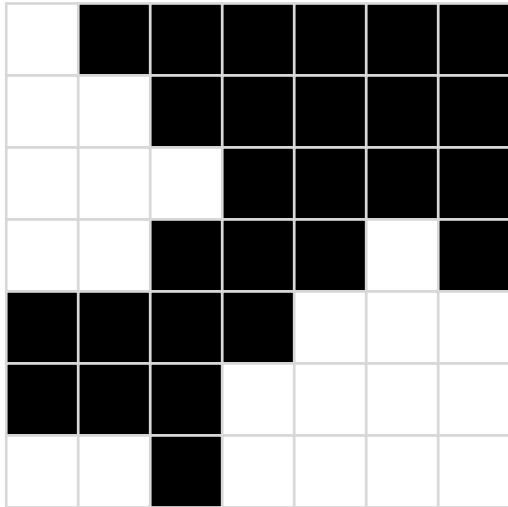
[y=152, x=92] = 0

No matter how they are displayed



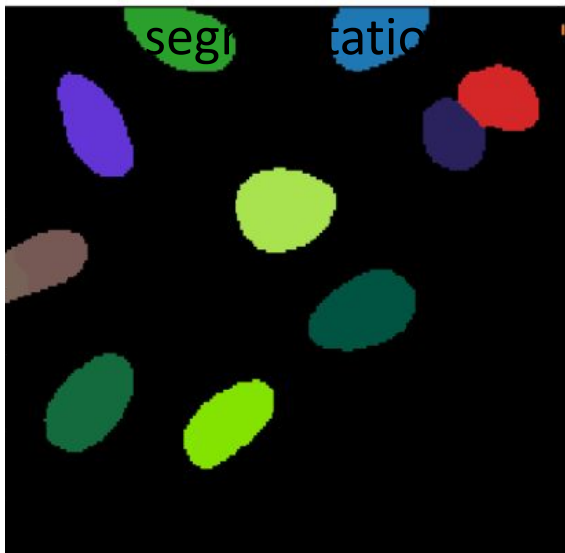
Connected component labelling

- In order to allow the computer differentiating objects, connected component analysis (CCA) is used to mark pixels belonging to different objects with different numbers
- Background pixels are marked with 0.
- The maximum intensity of a labelled map corresponds to the number of objects.



Terminology

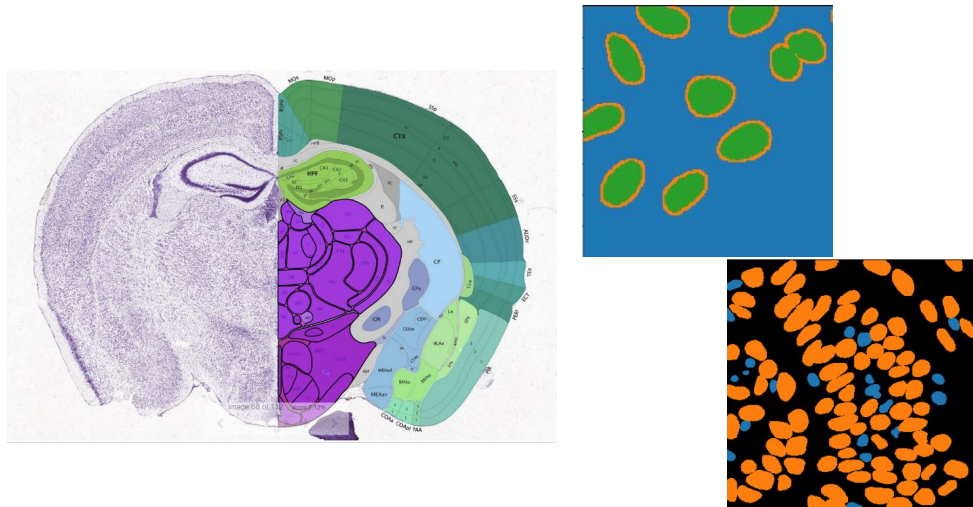
Instance



Instances:

- Cells, nuclei, cats, dogs, cars, trees

Semantic segmentation

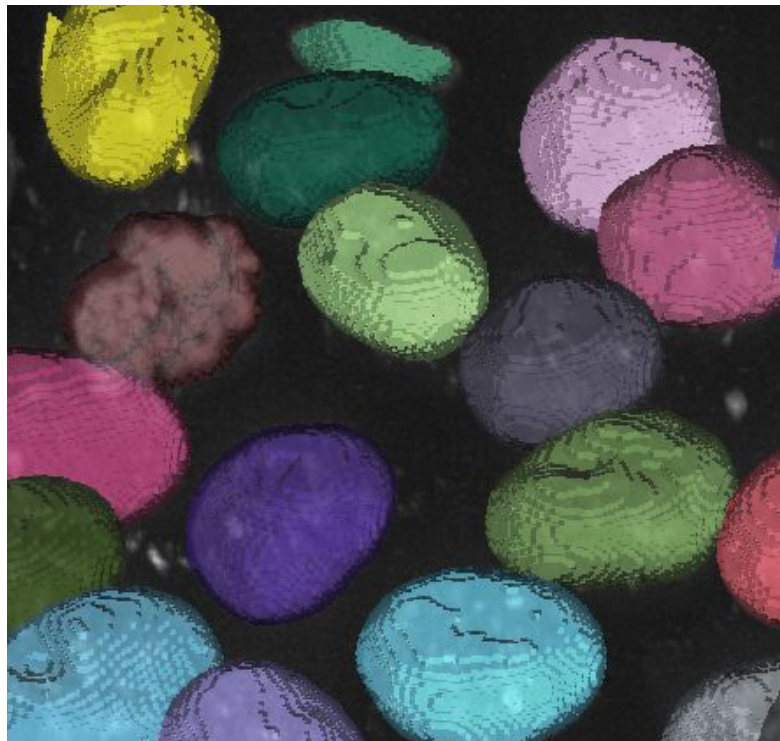


Regions:

- Anatomical, geographical
- All pixels belonging to the same type of object have the same value

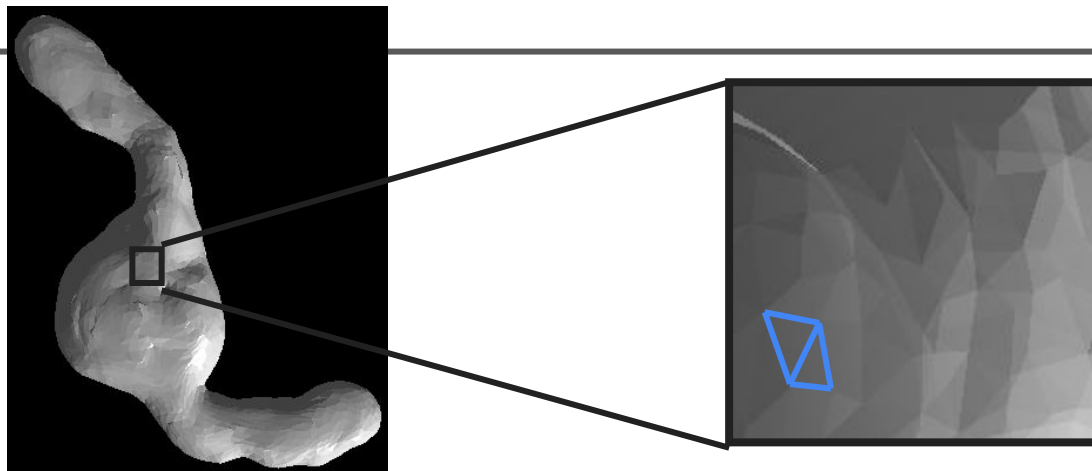
Motivation: Surface reconstruction

Pixel and voxel borders introduce artifacts, potentially problematic for measurements, e.g. surface area



Surface meshes

Points on a surfaces connected by triangles forma a surface mesh



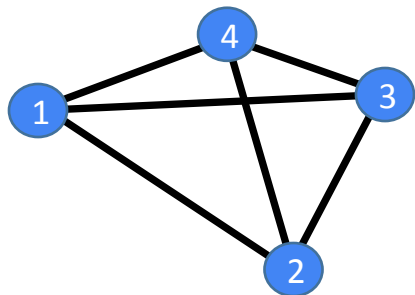
“Vertices” / points

Point x	Point y	Point z
x_1	y_1	z_1
x_2	y_2	z_2
x_3	y_3	z_3
x_4	y_4	z_4
...

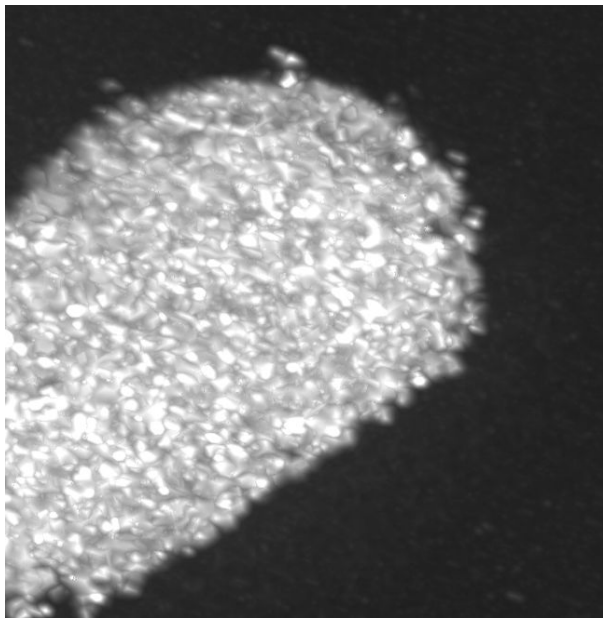
“Faces” / Triangles

Point 1	Point 2	Point 3
1	2	3
1	2	4
2	3	4
1	3	4

+



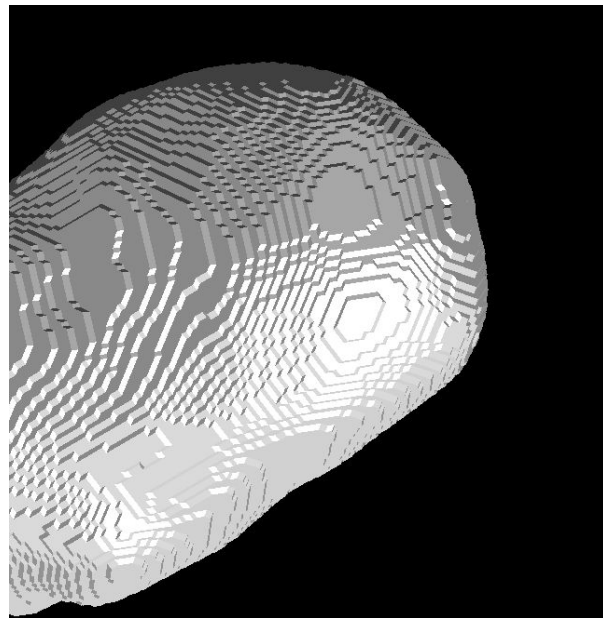
Surface reconstruction



3D image of nuclei



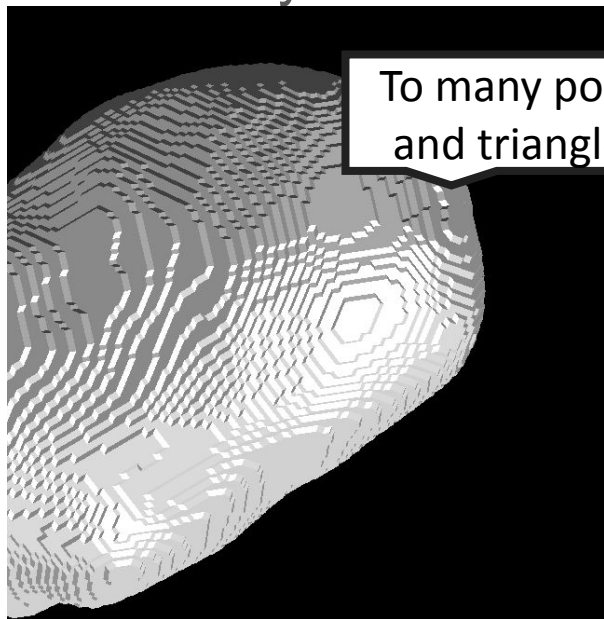
Gaussian filtered



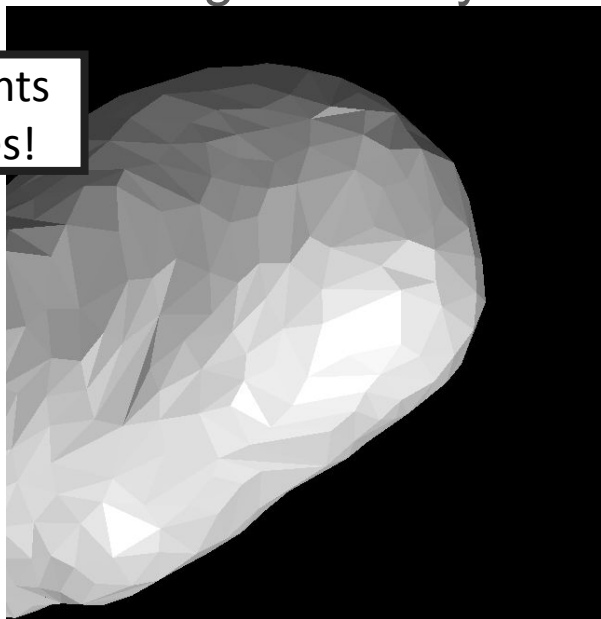
Binary 3D image
(visualized as surface mesh)

Surface post-processing

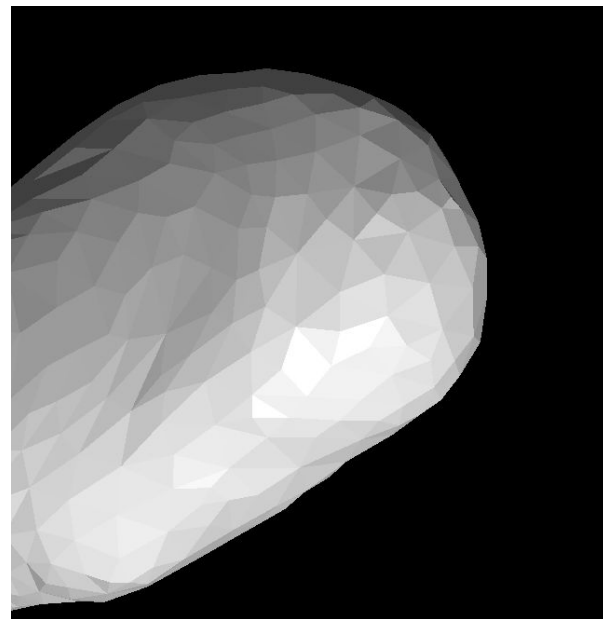
Necessary to better match biological reality.



Marching cubes result



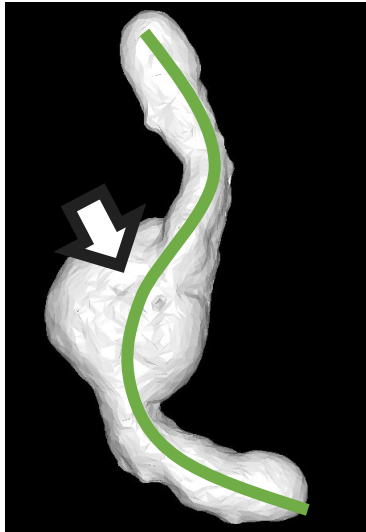
Simplified mesh
(less points, locally
averaged)



Smoothed mesh
(position locally planarized)

Surface post-processing

- Every processing step has consequences errors of later measurements
- Depends on desired measurement



Surface mesh



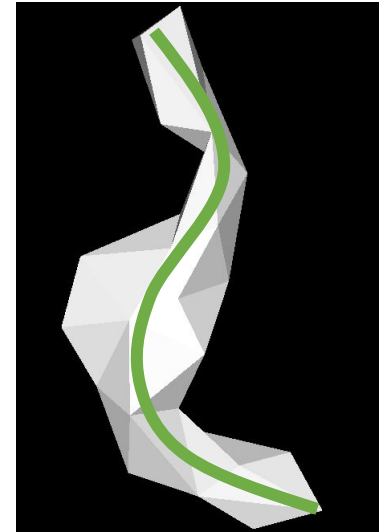
Simplified by factor 0.5

Number of small
concave regions



Simplified by factor 0.05

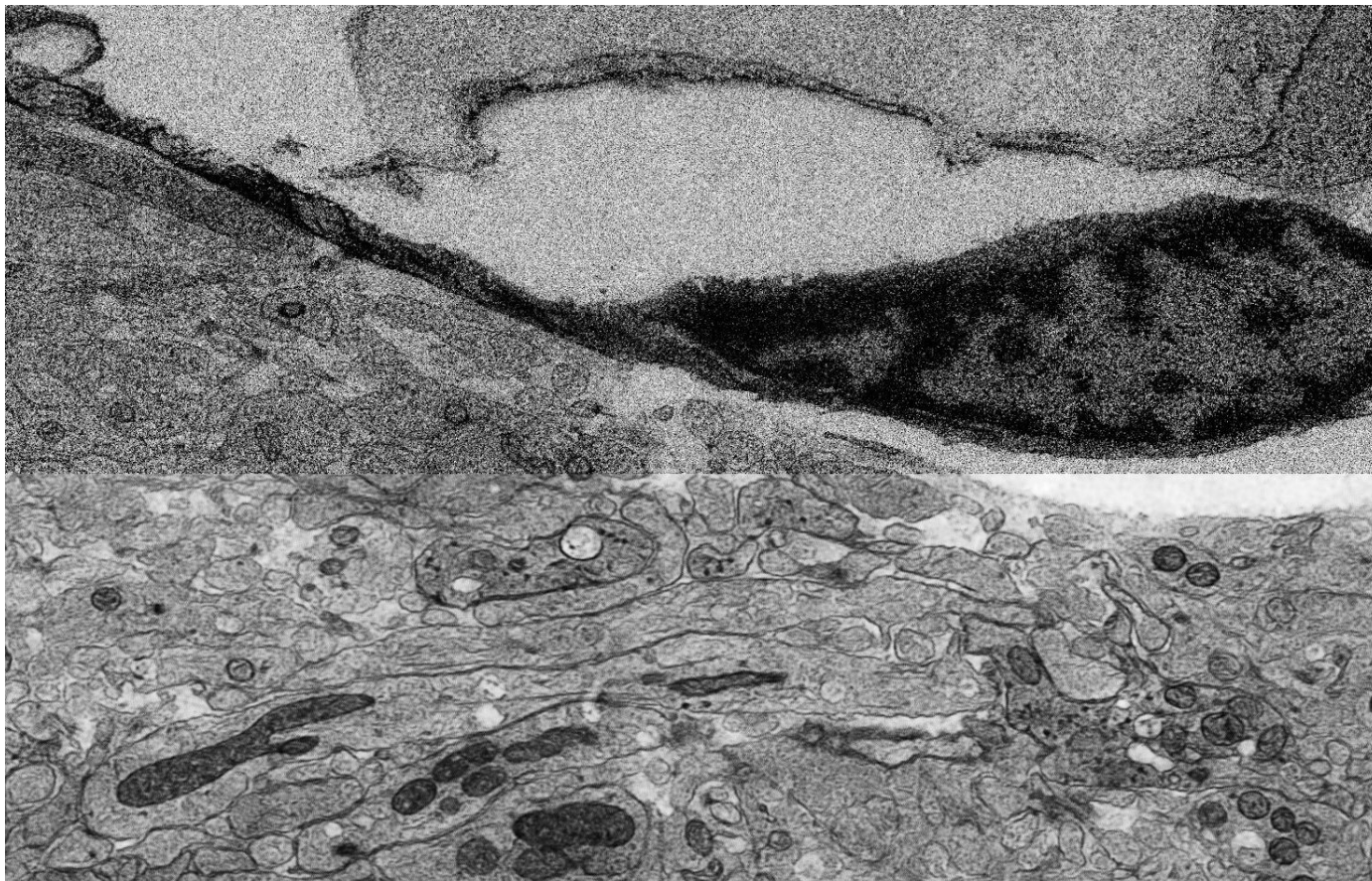
Total length



Simplified by factor 0.01

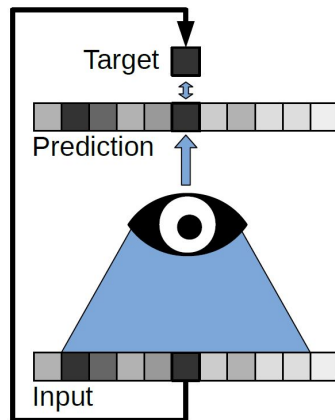
Bonus part

Deep Learning Denoising - Noise2Void

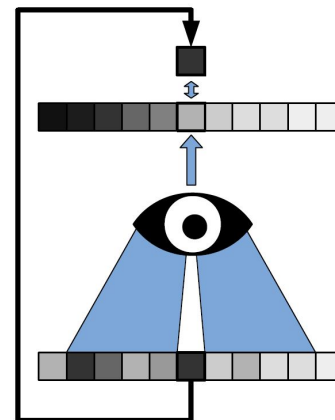


Deep Learning Denoising - Noise2Void

- Assumption: noise changes but image information remains.
 - The signal is statistically dependent on itself in the image.
 - Noise is statistically independent of the signal.
- Creating blind spots in the training image **(b)** will help the model recognize dependencies in the image.
- **Limitation:** Noise can be dependent based on digitalisation!



(a)

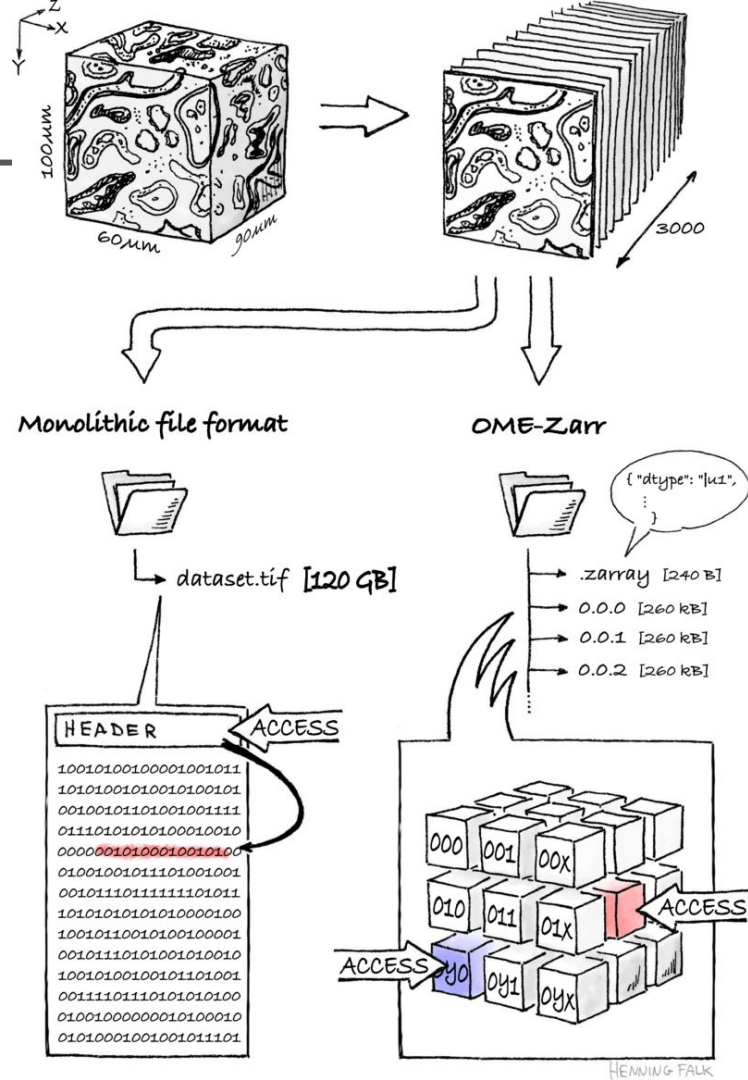


(b)

Zarr file format is structured into small, 3-dimensional chunks that can be directly accessed from the Zarr-directory. Hence, only the part of interest needs to be loaded instead of the full binary file.

This feature allows data to be accessed in sufficiently small pieces so that streaming the data of interest over a network instead of transferring the whole file becomes possible.

Falk, H. zarr-developers/zarr-illustrations-falk-2022 | Zenodo, 2022.
URL <https://doi.org/10.5281/zenodo.7037367>



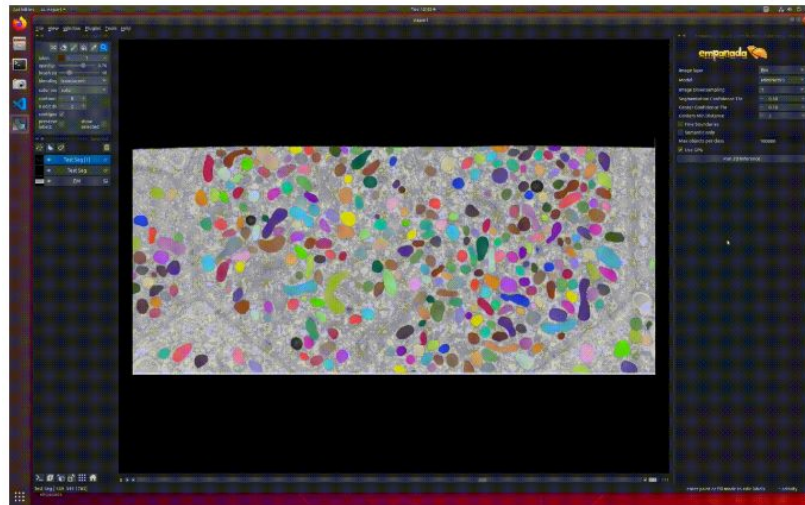
Visualize, share, and annotate your large 3D images online

The screenshot displays a web-based 3D visualization tool interface. The main window is divided into several panels:

- Top Panel:** Contains navigation icons (home, search, list, refresh, save), a menu dropdown, a coordinate system (2651, 1595, 1846), and various tool icons for manipulation and viewing.
- Left Panel (Layers/Settings):**
 - color:** Includes a histogram, opacity slider (set to 100), gamma correction slider (set to 1.00), and a color selection tool.
 - predictions, segmentation, Volume:** Each has a toggle switch.
 - Skeleton:** Has a toggle switch and several sliders: Node Radius (set to 1.00), Particle Size (set to 5), and Clipping Distance (set to 50.00). It also includes an Override Node Radius toggle (checked), and Auto-center Nodes and Highlight Commented Nodes toggles (unchecked).
 - Buttons:** A button labeled "+ Add Volume Annotation Layer" is at the bottom.
- Center Panel:** Shows a 3D view of a grayscale image with a blue box overlaid containing the text "SKELETON ANNOTATIONS". Below this, three 2D slices are visible: XY (top-left), YZ (top-right), and XZ (bottom-left). A blue skeleton is visible in the 3D view and the YZ slice.
- Right Panel (Info/Skeleton):**
 - Search and navigation icons.
 - A breadcrumb path: "explorative_2023-03-03_Lisa_Si".
 - A tree view under "Root" listing numerous items, each with a colored circle and a checkmark. The items are labeled with IDs and names like "explorative_2023-03-01_Lisa_f" and "explorative_2023-03-03_Lisa_f".
- Bottom Right:** A 3D view of the skeleton, showing a blue line with nodes. A legend below it shows "3D" with colored circles for XY (red), YZ (blue), and XZ (green). The URL "weblum.webknossos.org" is displayed at the bottom right.

empanada-napari

The empanada-napari plugin is built to democratize deep learning image segmentation for researchers in electron microscopy (EM). It ships with MitoNet, a generalist model for the instance segmentation of mitochondria. There are also tools to quickly build and annotate training datasets, train generic panoptic segmentation models, finetune existing models, and scalably run inference on 2D or 3D data. To make segmentation model training faster and more robust, CEM pre-trained weights are used by default. These weights were trained using an unsupervised learning algorithm on over 1.5 million EM images from hundreds of unique EM datasets making them remarkably general.



References

- Haase, R. (2021). Quantitative Bio-image Analysis. F1000 Research Limited. <http://doi.org/10.7490/F1000RESEARCH.1118789.1>
- Haase, R. (2023). Quantitative Bio-image Data Analysis in the age of Machine Learning. F1000 Research Limited. <http://doi.org/10.7490/F1000RESEARCH.1119447.1>
- Haase, R. (2023). Introduction to bio-image analysis I. F1000 Research Limited. <http://doi.org/10.7490/F1000RESEARCH.1119427.1>
- Haase, R. (2021). Interactive Data Visualization 101 with Fiji & Friends. F1000 Research Limited. <http://doi.org/10.7490/F1000RESEARCH.1118801.1>

Biolmage Analysis
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