

Katerina Mrazova

Staining Strategies of Biological Samples Prepared for Volume Microscopy

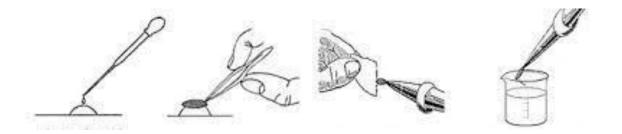
Summary



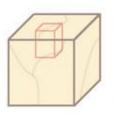
- On section vs. En bloc staining
- Staining reagents
 - → osmium tetroxide
 - → thiokarbohydrazide
 - → uranyl acetate
 - → lead aspartate
- Mostly used methods
 - \rightarrow OTO
 - \rightarrow rOTO
 - \rightarrow Hua
- Possible problems
- Alternative staining methods

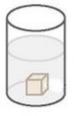
Staining strategies















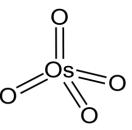




Staining reagents



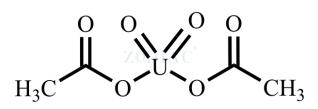
- Osmium tetroxide
 - → yellow crystals, highly oxidizing, volatile vapours
 - → reaction with organic compounds (unsaturated bonds of fatty acids)
 - → secondary fixation agent (membranes) as well as a contrasting agent
 - \rightarrow reduced osmium (+ K₃[Fe(CN)₆] / K₄[Fe(CN)₆])
- Thiokarbohydrazide
 - → white to pale grey crystals, toxic, light-sensitive
 - → very slightly soluble (0.5g/100g, 25°C)
 - → attachment to osmium bound in the tissue enabling second osmium binding



Staining reagents



- Uranyl acetate
 - → negative staining, on-section staining, en bloc staining since the 1960s
 - → highly toxic, mildly radioactive
 - → mostly reacts with nucleic acids and proteins
 - → subject to rising legal restrictions
 - → lanthanoids as a possible substitute
- Lead aspartate (Walton PbAsp, 1979)
 - → aspartic acid + lead nitrate
 - → lesser contaminations, lower pH than lead citrate
 - → toxic, challenging preparation



Conventional staining strategies



	protocol					
incubation steps	гото	ото	Hua			
1.	2% OsO4, 2.5% ferrocyanide, 0.15 M Cac, pH 7.4	2% OsO4, unbuffered	2% OsO4, 0.15 M Cac, pH 7.4			
	1.5 h @ rt	1.5 h @ rt	1.5 h @ rt			
	No wash					
2			2.5% ferrocyanide, 0.15 M Cac, pH 7.4			
			1.5 h @ rt			
	0.5 h wash in water x 2					
3	1% TCH, unbuffered	1% TCH, unbuffered	1% TCH, unbuffered			
	0.75 h @ 50 °C	0.75 h @ rt	0.75 h @ 40 °C			
	0.5 h wash in water x 2					
4	2% OsO4,	2% OsO4,	2% OsO4,			
	unbuffered 1.5 h @ rt	unbuffered 1.5 h @ rt	unbuffered 1.5 h @ rt			
	0.5 h wash in water x 2					
	1 % uranyl acetate, unbuffered	1 % uranyl acetate, unbuffered	1% uranyl acetate, unbuffered			
5	2 h @ 50 °C	2 h @ 50 °C	overnight @ 4 °C, 2 h @ 50 °C			
	0.5 h wash in water x 2					
6	Lead aspartate, pH 5.0	Lead aspartate, pH 5.0	Lead aspartate, pH 5.0			
	2 h @ 50 °C	2 h @ 50 °C	2 h @ 50 °C			
		0.5 h wash in water x 2				
	dehydration, infiltration and embedding					

OTO, Seligman, 1966

ISI CAS

- Firstly published to enhance the contrast of osmicated sections on grids
- Variations of the protocol used for en bloc staining
- Procedure
 - \rightarrow glutaraldehyde (2,5% in buffer, RT/4 °C, 4h)
 - \rightarrow washing buffer (3x15min)
 - \rightarrow OsO₄ (2% in buffer, RT, 1,5h)
 - \rightarrow washing buffer (3x15min)
 - → thiocarbohydrazide (1% in water, 50°C, 1h)
 - \rightarrow washing water (3x15min)
 - \rightarrow OsO₄ (1% in water, RT, 1h)
 - \rightarrow washing water (3x15min)
 - \rightarrow uranyl acetate (1% in water, 50°C, 2h)
 - \rightarrow washing water (3x15min)
 - → Walton lead aspartate (50°C, 2h)
 - \rightarrow washing water (3x15min)
 - → acetone (30% » 50% » 70% » 80% » 90% » 95% » 100%, RT, 15min)
 - → epon (in acetone, 1:2 » 1:1 » 2:1 » 2x pure resin, RT, 1h, last overnight, curing 60°C 48h)

rOTO, Willingham, 1983



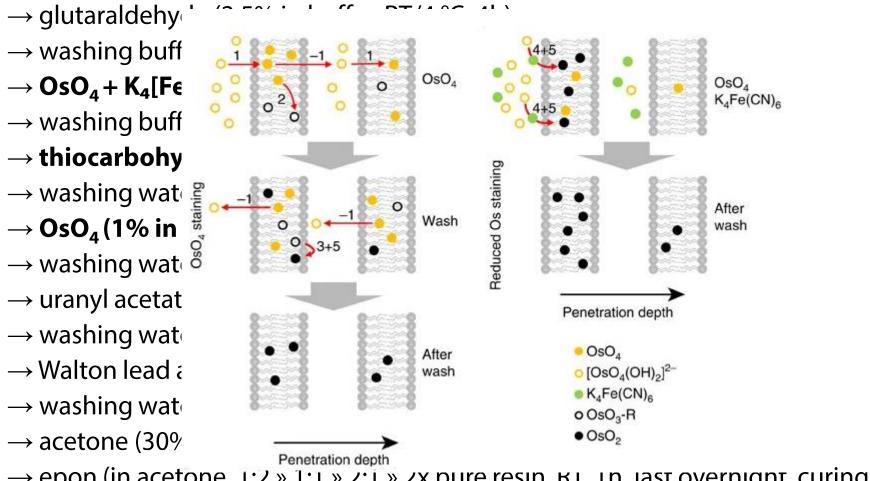
- Improvement of fixation/staining of lipidic structures and membranes before EtOH dehydration
- Procedure
 - \rightarrow glutaraldehyde (2,5% in buffer, RT/4 °C, 4h)
 - \rightarrow washing buffer (3x15min)
 - \rightarrow OsO₄ + K₄[Fe(CN)₆] (2%+2,5% in buffer, RT, 1,5h)
 - \rightarrow washing buffer (3x15min)
 - → thiocarbohydrazide (1% in water, 50°C, 1h)
 - \rightarrow washing water (3x15min)
 - \rightarrow OsO₄ (1% in water, RT, 1h)
 - \rightarrow washing water (3x15min)
 - \rightarrow uranyl acetate (1% in water, 50°C, 2h)
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Improvement of fixation/staining of lipidic structures and membranes before EtOH dehydration

Procedure



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Hua, 2015

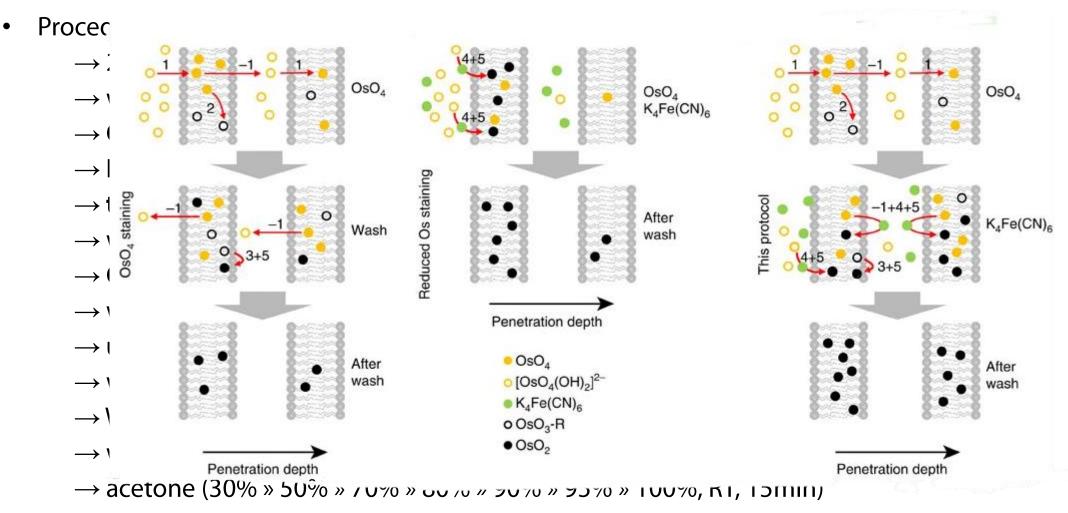


- Main changes in Os and U steps to achieve high-contrast staining throughout large tissue blocks
- Procedure
 - \rightarrow 2,5% glutaraldehyde (in buffer, RT/4 °C, 4h)
 - \rightarrow washing buffer (3x15min)
 - \rightarrow OsO₄ (2% in buffer, RT, 1,5h)
 - \rightarrow K₄[Fe(CN)₆] (2,5% in buffer, RT, 1,5h)
 - → thiocarbohydrazide (1% in water, 40°C, 45min)
 - \rightarrow washing water (3x15min)
 - \rightarrow OsO₄ (2% in water, RT, 1,5h)
 - → washing water (3x15min)
 - → uranyl acetate (1% in water, 4°C overnight, 50°C 2h)
 - \rightarrow washing water (3x15min)
 - \rightarrow Walton lead aspartate (50°C, 2h)
 - \rightarrow washing water (3x15min)
 - → acetone (30% » 50% » 70% » 80% » 90% » 95% » 100%, RT, 15min)
 - → epon (in acetone, 1:2 » 1:1 » 2:1 » 2x pure resin, RT, 1h, last overnight, curing 60°C 48h)

Hua, 2015



Main changes in Os and U steps to achieve high-contrast staining throughout large tissue blocks

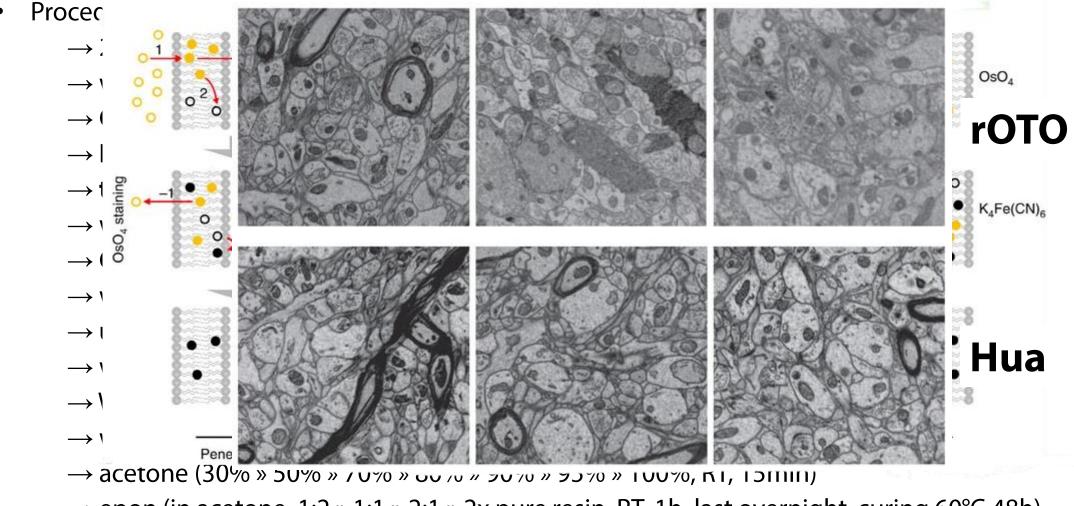


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Hua, 2015



Main changes in Os and U steps to achieve high-contrast staining throughout large tissue blocks



 \rightarrow epon (in acetone, 1:2 » 1:1 » 2:1 » 2x pure resin, RT, 1h, last overnight, curing 60°C 48h)

Issues



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Issues



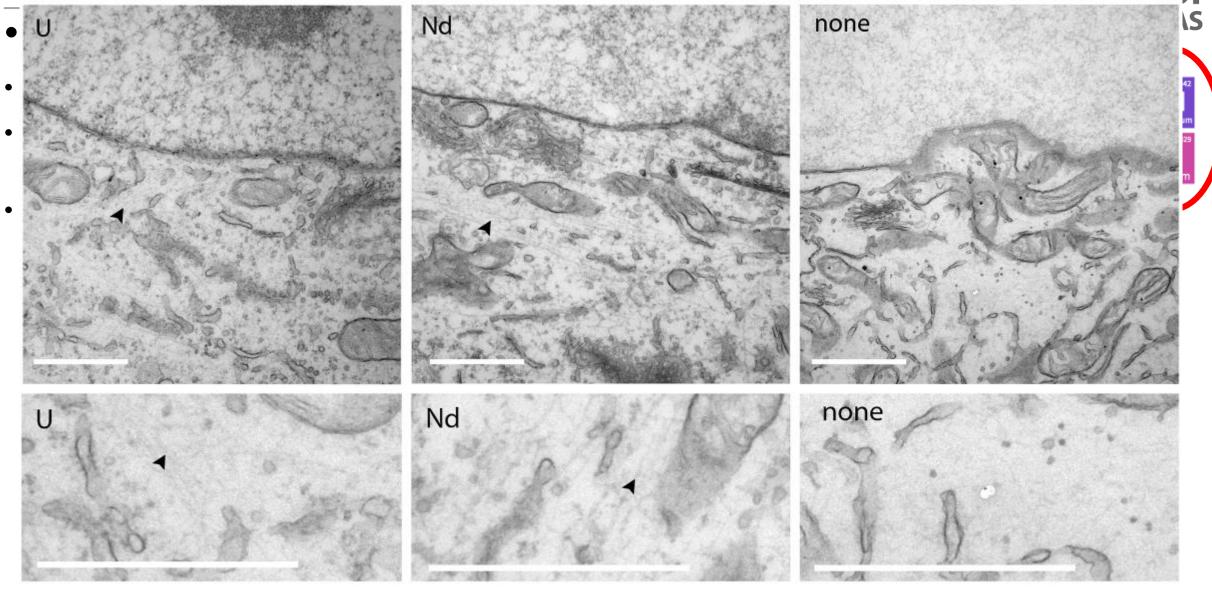
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- Kuipers, 2020
- Use of neodymium acetate as uranyl substitute
- Simillar chemical properties due to the position in the table of elements therefore assumption → very similar in binding to tissue
- Procedure
 - → standard fixation and postfixation by osmium
 - \rightarrow 4% NdAc 30 / 60 / 120 min at RT
 - → dehydration and resin embedding

*Lanthanoids

**Actinoids





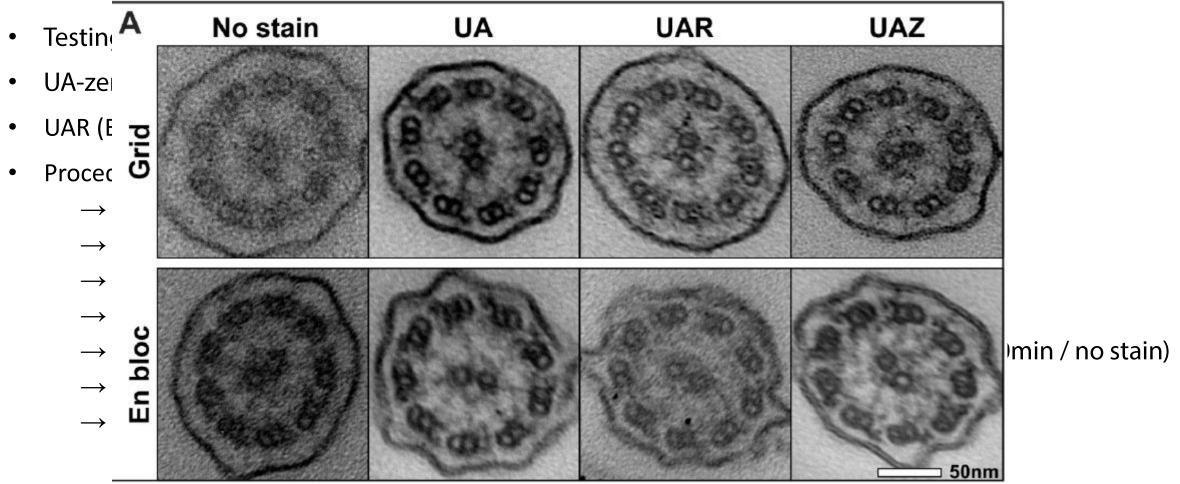
• Pinto, 2021



- Testing commercially available uranyl-less staining agents on cilia
- UA-zero (Agar Scientific) → ytterbium chloride +phosphothungstid acid
- UAR (EMS) → samarium and gadolinium triacetate
- Procedure
 - → glutaraldehyde (2,5% in buffer, 4°C, overnight)
 - \rightarrow wash (buffer)
 - \rightarrow OsO₄ (1% in water, RT, 1h)
 - \rightarrow wash (water)
 - → UA/Ua zero/UAR/no stain (1% in water 30min / no dilution 30min / 1:4 in water 30min / no stain)
 - → ethanol (50% » 70% » 90% » 100%)
 - → propylene oxide + resin



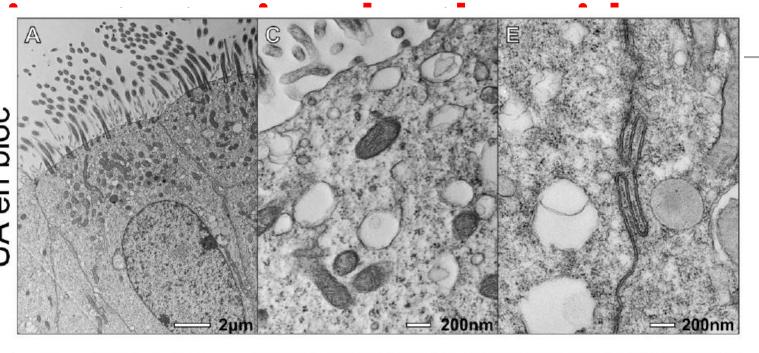
• Pinto, 2021

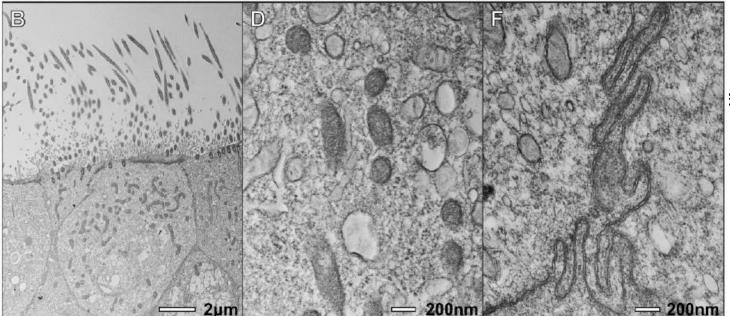


Alterna

- Pinto, 2
- Testing com
- UA-zero (Ag
- Procedure
 - → gluta
 - \rightarrow wash
 - \rightarrow OsO₄ (
 - \rightarrow wash
 - \rightarrow UA/U $\stackrel{\circ}{\sim}$ $\stackrel{\circ}{\rightarrow}$ ethan $\stackrel{\circ}{\sim}$

 - → propy **ō**







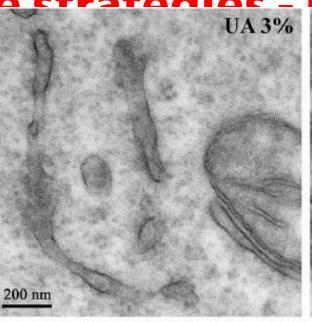
iter 30min / no stain)

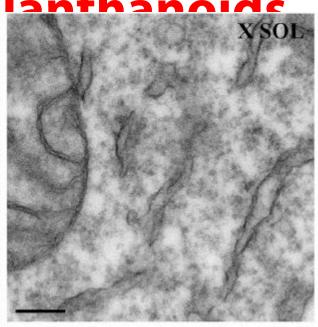
ISI CAS

- Moscardini, 2020
- Use of ytterbium chloride and phosphotungstic acid (PTA) as an alternative stain
- Commercially available as UA zero (Agar Scientific)
- For negative staining, on-section staining, en bloc staining
- Ytterbium high electron scattering power, PTA previously proven to enhance Uac staining

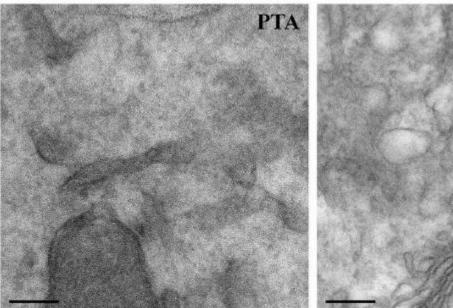
Moscardini,

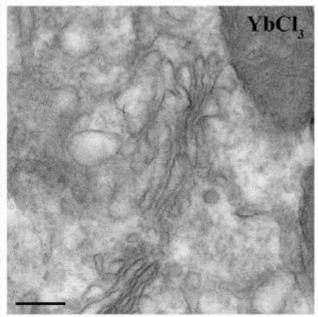
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- Commercially avail
- For negative staini
- Ytterbium high ele









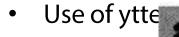


ISI CAS

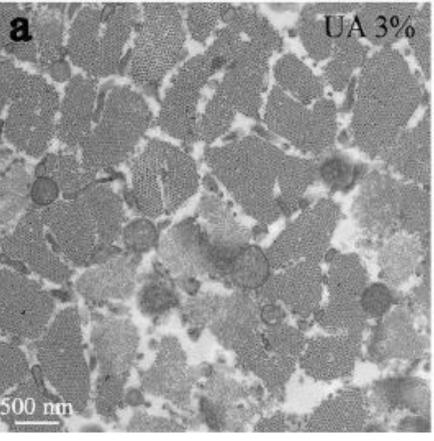
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- Commercially available as UA zero (Agar Scientific)
- For negative staining, on-section staining, en bloc staining
- Ytterbium high electron scattering power, PTA previously proven to enhance Uac staining
- Procedure
 - → glutaraldehyde (2% in buffer, 4°C, overnight)
 - \rightarrow OsO₄ + K₃[Fe(CN)₆] (1% +1% in buffer)
 - \rightarrow washing
 - \rightarrow optimized X Solution (ratio 15 YbCl : 1 PTA), PTA 3.2 mM, YbCl₃ 48 mM alone and UA 3% (1h)
 - → dehydration, resin embedding

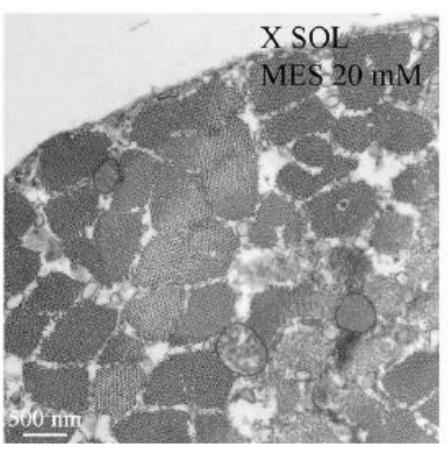


• Moscardini, 2020



- Commerci
- For negating
- Ytterbium
- Procedure
 - → glut
 - \rightarrow OsO
 - \rightarrow was
 - → opti
 - \rightarrow dehy





% (1h)

Alternative strategies - lanthanoids ISI CAS Uranyless Mosc Use of yt Commer For nega Ytterbiui Procedu $\rightarrow gli$ UAR \rightarrow Os $\longrightarrow \mathsf{W}\check{\epsilon}$ (1h) \rightarrow op \rightarrow de



Thank you for your attention.